Circulating androgen levels are not affected by the administration of vaginal micronized progesterone for withdrawal bleeding in patients with polycystic ovary syndrome

Carlos Dosouto MD1, Jorge Rodríguez-Purata MD1, Beatriz Álvaro PhD2, Sofia Gaggiotti-Marre MD1, Ignacio Rodríguez MD2, Francisca Martínez PhD1


BACKGROUND: Polycystic ovary syndrome (PCOS) is the most common endocrine disorder in women of reproductive age, with hyperandrogenemia as one of its main components. Several medications have been described to properly induce withdrawal bleeding, unfortunately, most of them significantly suppress ovarian function and lower circulating androgen levels with a subsequent diagnosis bias. The present study aims to determine whether the use of vaginal micronized progesterone (VMP) to induce withdrawal bleeding in women with PCOS alters circulating androgen parameters.

METHODS: This is a prospective clinical trial, performed at an academic medical center. Fifteen reproductive-aged women with PCOS were studied. Blood samples were collected at baseline (Sample 1) and between the 3rd-5th day of withdrawal after 7 days of VMP (100 mg every 12 h) (Sample 2). Total testosterone (TT), free androgen index (FAI), sex hormone binding globulin (SHBG), dehydroepiandrosterone sulfate (DHEAS), androstenedione (A4) and 17-hydroxyprogesterone (17-OHP) levels were determined in the blood samples.

RESULTS: All 15 patients fulfilled the Rotterdam criteria for PCOS. Two patients presented spontaneous ovulation confirmed in Sample 1 and were not included in the analysis. Mean age was 29.3 ± 4.9 years and mean body mass index was 26.9 ± 6.7 kg/m². The mean values of TT, FAI, SHBG, DHEAS, A4 and 17-OHP did not change after VMP administration.

CONCLUSION: Withdrawal bleeding with VMP in PCOS patients does not significantly alter circulating androgen levels compared to baseline and can thus be used to time blood sampling in these patients.

Key Words: Polycystic ovarian syndrome, androgens, progesterone, withdrawal, hyperandrogenism

ABBREVIATIONS: PCOS Polycystic ovary syndrome; 17-OHP 17-hydroxyprogesterone; MPA Medroxyprogesterone; MP Micronized progesterone; VMP Vaginal micronized progesterone; OMP Oral micronized progesterone; TT Total testosterone; FAI Free androgen index; SHBG Sex hormone binding globulin; DHEAS Dehydroepiandrosterone sulfate; A4 Androstenedione; RIA Radioimmuno assay; FSH Follicle stimulating hormone; LH Luteinizing hormone; E2 Estradiol; BMI Body mass index

Polycystic ovarian syndrome (PCOS) is the most common endocrine disorder in women of reproductive age, affecting 7% of the female population (1). While the etiology of PCOS is complex and not completely understood, the syndrome is characterized by oligo/anovulation with polycystic ovaries and increased androgen secretion resulting in acne and hirsutism (2). Hormonal evaluation of women who are suspected of having PCOS involves the measurement of basal levels of androgens and 17-hydroxyprogesterone (17-OHP), which are generally used to establish the presence of hyperandrogenemia and exclude other diagnoses with similar clinical signs, as nonclassic congenital adrenal hyperplasia (1). In general, these levels are obtained during the early follicular phase to maintain sampling uniformity and avoid spurious increases due to corpus luteum function. However, because most hyperandrogenic patients are oligo/amenorrheic, it is frequently necessary to administer a progestogen to induce withdrawal bleeding and properly time the blood sampling.

Several medications have been described to properly induce withdrawal bleeding (3), with medroxyprogesterone acetate (MPA) being the most widely used. However, synthetic compounds as MPA do not replicate precisely the constellation of biologic activities of the parent hormone (4). Previous reports have suggested that the use of MPA in women with PCOS results in a temporary, albeit clinically relevant, suppression in ovarian function (5) and circulating androgen levels (6,7), in addition to several adverse side effects (3). All comparative studies to date conclude that the side effects of synthetic progestins can be minimized or eliminated through the use of micronized progesterone (MP) (8-10) and consequently result in reduced hormonal suppression (11).

The putative advantage of using MP over MPA is that the former is bioidentical to the body’s own progesterone produced by the corpus luteum. Recent evidence has shown that MP displays a favorable action on blood vessels and on the brain, which may not be true for some synthetic progestins (12). Furthermore, MPA, possibly owing to its glucocorticoid activity, may counteract some of the beneficial effects of estradiol, which is not the case with MP. Moreover, there is evidence that synthetic progestins have a teratogenic effect when administered during the first 4 months of pregnancy (10). Woods et al previously investigated if oral MPA used to induce withdrawal bleeding in women suspected of having PCOS alters circulating androgen levels (11). The authors concluded that the administration of oral MPA did not significantly alter circulating androgen or 17-OHP levels, and could be used to time blood sampling in these patients. Nonetheless, it has been previously described the dose, duration, and route of administration are critical to optimization of the treatment regimen (13).

In this study, it is hypothesized that the administration of natural progesterone vaginally, which will avoid hepatic first pass, may result in significantly less hormonal suppression. The authors test this hypothesis by prospectively determining the effect of vaginal MP (VMP) administered for the induction of withdrawal bleeding, on the circulating androgen and 17-OHP levels in women with PCOS.

METHODS

Settings and ethics

All patients who came into consult of the doctor in charge of the study and responsible for the Reproductive Endocrinology Unit at Hospital Universitario Dexeus and fulfilled the inclusion criteria, were offered to participate. The study was approved by the Institutional Review Board, and all participants were provided written informed consent.

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Hormonal analysis

The levels of total testosterone (TT), free androgen index (FAI), sex hormone-binding globulin (SHBG), dehydroepiandrosterone sulfate (DHEAS), androstenedione (A4) and 17-OHP were measured by standard radioimmunoassay (RIA). Follicle stimulating hormone (FSH), luteinizing hormone (LH) and estradiol (E2) levels were also evaluated to assess eventual spontaneous ovulation. The following commercially available kits were used: Gamma Counter Wizard by Perkin Elmer for 17-OHP, A4 and FAI; Beckman Coulter (Access) for DHEAS, TT, SHBG, FSH, LH and E2. The intra-assay coefficient of variation for all steroids did not exceed 6%, except for estradiol, which was 12%.

Collection of samples

All patients were sampled on the day of the first visit at the outpatient clinic (Sample 1). Sample 2 was obtained between the 3rd and 5th day of induced withdrawal. Blood samples 2 were drawn in the morning between 8 and 11 am, after each patient had been fasting for at least 8 h and centrifuged immediately and serum was stored at -80ºC until it was assayed.

TABLE 1
Group differences in hormone values between baseline and after VMP administration

<table>
<thead>
<tr>
<th></th>
<th>Before VMP (n=13)</th>
<th>After VMP (n=13)</th>
<th>p Value</th>
<th>Normal value</th>
</tr>
</thead>
<tbody>
<tr>
<td>TT (ng/mL)</td>
<td>0.6 ± 0.1</td>
<td>0.5 ± 0.1</td>
<td>0.39</td>
<td>0.15-0.7</td>
</tr>
<tr>
<td>FAI (%)</td>
<td>5.2 ± 3.2</td>
<td>4.7 ± 2.7</td>
<td>0.16</td>
<td>0.6-10.6</td>
</tr>
<tr>
<td>A4 (ng/mL)</td>
<td>2.5 ± 1.0</td>
<td>2.8 ± 1.3</td>
<td>0.53</td>
<td>0.4-3.4</td>
</tr>
<tr>
<td>DHEAS (mcmol/L)</td>
<td>2.2 ± 1.9</td>
<td>2.3 ± 2.1</td>
<td>0.46</td>
<td>0.92-11.67</td>
</tr>
<tr>
<td>17-OHP (ng/mL)</td>
<td>0.6 ± 0.2</td>
<td>0.7 ± 0.5</td>
<td>0.89</td>
<td>0.11-1.8</td>
</tr>
<tr>
<td>SHBG (nmol/L)</td>
<td>52.3 ± 22.2</td>
<td>54.3 ± 26.7</td>
<td>0.54</td>
<td>18-144</td>
</tr>
<tr>
<td>FSH (UI/L)</td>
<td>5.4 ± 1.2</td>
<td>5.0 ± 1.6</td>
<td>0.40</td>
<td>1.4-9.9</td>
</tr>
<tr>
<td>LH (UI/L)</td>
<td>11.8 ± 7.9</td>
<td>8.1 ± 4.4</td>
<td>0.06</td>
<td>1.7-15</td>
</tr>
<tr>
<td>TT (ng/mL)</td>
<td>0.6 ± 0.1</td>
<td>0.5 ± 0.1</td>
<td>0.39</td>
<td>0.15-0.7</td>
</tr>
<tr>
<td>FAI (%)</td>
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<td>4.7 ± 2.7</td>
<td>0.16</td>
<td>0.6-10.6</td>
</tr>
<tr>
<td>A4 (ng/mL)</td>
<td>2.5 ± 1.0</td>
<td>2.8 ± 1.3</td>
<td>0.53</td>
<td>0.4-3.4</td>
</tr>
<tr>
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</tr>
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<td>0.40</td>
<td>1.4-9.9</td>
</tr>
</tbody>
</table>

TABLE 2
Individual values for each patient before (1) and after (2) VMP administration. Abnormal values are highlighted in bold

<table>
<thead>
<tr>
<th>n</th>
<th>TT1 (ng/mL)</th>
<th>TT2 (ng/mL)</th>
<th>FAI1 (%)</th>
<th>FAI2 (%)</th>
<th>A41 (ng/mL)</th>
<th>A42 (ng/mL)</th>
<th>DHEAS1 (mcmol/L)</th>
<th>DHEAS2 (mcmol/L)</th>
<th>17-OHP1 (ng/mL)</th>
<th>17-OHP2 (ng/mL)</th>
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</thead>
<tbody>
<tr>
<td>1</td>
<td>1</td>
<td>0.9</td>
<td>8.23</td>
<td>6.54</td>
<td>4.1</td>
<td>3.4</td>
<td>1.05</td>
<td>1.01</td>
<td>1.03</td>
<td>0.75</td>
</tr>
<tr>
<td>2</td>
<td>0.4</td>
<td>0.4</td>
<td>4.15</td>
<td>1.67</td>
<td>1.2</td>
<td>1.6</td>
<td>1.62</td>
<td>1.28</td>
<td>0.4</td>
<td>1.5</td>
</tr>
<tr>
<td>3</td>
<td>0.6</td>
<td>0.5</td>
<td>3.13</td>
<td>2.9</td>
<td>2.3</td>
<td>2.2</td>
<td>0.9</td>
<td>0.91</td>
<td>0.82</td>
<td>0.69</td>
</tr>
<tr>
<td>4</td>
<td>0.8</td>
<td>0.9</td>
<td>6</td>
<td>4.92</td>
<td>4.2</td>
<td>5.3</td>
<td>2.27</td>
<td>6.41</td>
<td>1.25</td>
<td>2.05</td>
</tr>
<tr>
<td>5</td>
<td>0.6</td>
<td>0.4</td>
<td>2.27</td>
<td>1.68</td>
<td>2.9</td>
<td>1.8</td>
<td>1.93</td>
<td>1.51</td>
<td>0.73</td>
<td>0.34</td>
</tr>
<tr>
<td>6</td>
<td>0.5</td>
<td>0.6</td>
<td>4.24</td>
<td>4.78</td>
<td>2.7</td>
<td>5</td>
<td>2.43</td>
<td>1.75</td>
<td>0.7</td>
<td>0.99</td>
</tr>
<tr>
<td>7</td>
<td>0.7</td>
<td>0.6</td>
<td>12.28</td>
<td>10.79</td>
<td>1.8</td>
<td>2.9</td>
<td>2.4</td>
<td>3.35</td>
<td>0.36</td>
<td>0.46</td>
</tr>
<tr>
<td>8</td>
<td>0.4</td>
<td>0.5</td>
<td>4.48</td>
<td>8.27</td>
<td>1.5</td>
<td>1.4</td>
<td>1.71</td>
<td>1.48</td>
<td>0.53</td>
<td>0.35</td>
</tr>
<tr>
<td>9</td>
<td>0.5</td>
<td>0.9</td>
<td>3.13</td>
<td>4.66</td>
<td>2.3</td>
<td>2.1</td>
<td>1.34</td>
<td>1.62</td>
<td>0.85</td>
<td>0.76</td>
</tr>
<tr>
<td>10</td>
<td>0.4</td>
<td>0.5</td>
<td>1.67</td>
<td>1.61</td>
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<td>2.1</td>
<td>1.6</td>
<td>1.04</td>
<td>0.5</td>
<td>0.7</td>
</tr>
<tr>
<td>11</td>
<td>0.6</td>
<td>0.4</td>
<td>3.44</td>
<td>2.66</td>
<td>2.4</td>
<td>2.4</td>
<td>2.49</td>
<td>2.37</td>
<td>0.93</td>
<td>0.31</td>
</tr>
<tr>
<td>12</td>
<td>0.7</td>
<td>0.4</td>
<td>11.52</td>
<td>4.93</td>
<td>3.2</td>
<td>1.9</td>
<td>0.87</td>
<td>0.77</td>
<td>0.73</td>
<td>0.2</td>
</tr>
<tr>
<td>13</td>
<td>0.8</td>
<td>0.5</td>
<td>6.27</td>
<td>5.85</td>
<td>3.3</td>
<td>4.7</td>
<td>8.98</td>
<td>7.3</td>
<td>0.46</td>
<td>0.48</td>
</tr>
</tbody>
</table>
All hormonal parameters were compared before and after the administration of 200 mg of VMP (Utrogestan®; Solvay-Pharma, Suresnes, France) (100 mg every 12 h) for a 7 day course.

**Statistical analysis**

All statistical analyses were performed using the IBM® SPSS® Statistics v22.0 application. Data are expressed as means ± SD. Difference between the two groups was compared by Wilcoxon Rank Test for paired data. When variables were categorized (normal or abnormal) by our reference laboratory, the MacNemar test was used.

**Main outcome measures**

The main outcome measure was any differences on androgen level before and after VMP administration. As a secondary outcome, we evaluated whether this regimen affects the diagnosis of analytical hyperandrogenism.

**RESULTS**

A total of fifteen Caucasian reproductive-aged women with PCOS were recruited. All fifteen patients fulfilled the Rotterdam criteria for diagnosis of PCOS. Of these, 13 patients were anovulatory and two patients presented spontaneous ovulation, confirmed by Sample 1 and were not included in the analysis. Ovulation was confirmed by a serum progesterone level >1.5 ng/mL. Their mean age was 29.3 ± 4.9 years and mean body mass index (BMI) was 26.9 ± 6.7 Kg/m². Baseline hormonal characteristics of the patients are given in Table 1.

The mean values of TT, FAI, SHBG, DHEAS, A4 and 17-OHP did not change with VMP administration, and neither did the mean values of FSH, LH and E2. There was a tendency to observe lower levels of LH during withdrawal bleeding in comparison with baseline values, albeit without significant differences (Table 1).

There were no significant differences when the diagnosis of analytical hyperandrogenism (normal or abnormal according to our reference laboratory) before and after the VMP regimen was compared (Table 2).

**DISCUSSION**

The clinical value of these studies is to help with the diagnosis of PCO, ensure withdrawal and better programming of blood hormonal tests.

The data from the present study indicate that in women with anovulatory PCOS, androgen hormonal profile (TT, FAI, SHBG, DHEAS, A4) and 17-OHP did not significantly differ at baseline and after VMP administration.

The induction of withdrawal bleeding with progestins has been considered a common procedure in several studies to compare hormonal values in follicular phase among patients with PCOS. The reported data on this topic is poor, unclear, with a small number of patients assessed, and they are not homogeneous in regard to PCOS Rotterdam criteria. In many cases, withdrawal is performed with different kinds of progestins (Depot Medroxyprogesterone or oral Micronized Progestrone), rendering it incomparable (11,15).

However, the usefulness of this test requires further investigation in PCOS, as the available data are reported with MPA (5,7,11,16-21) or oral MP and for a not well-established period of time (22). Previous studies (5-7,16), have shown that progestosterone administration could suppress pituitary LH secretion, ovarian function and androgen synthesis, suggesting that this could be a diagnostic bias (17).

We used vaginal administration for several reasons. On the one hand, this form of administration could cause fewer metabolic changes by avoiding hepatic first pass (15,23). In addition, it offers better tolerance (24), excellent absorption very close to the target organ; it avoids oral administration and is the alternative preferred by patients (23).

The progestin used in the present study (Utrogestan®) was synthesized from a precursor extracted from yam (Dioscorea sp.) and is chemically identical to ovarian progesterone (25,26). Its micronized formulation reportedly provides optimal progesterone bioavailability after vaginal administration, reaching its maximum level after 2 to 6 h and returning to baseline values within 24 h after the last dose. One dose every 12 h ensures the least variation between individuals (27). Thus, in our opinion, it could be an optimal regimen for the diagnostic assessment in PCOS patients, being useful to induce withdrawal bleeding without significantly changing circulating androgen levels. The fact that every patient is their own control is a strong point of the study, although our results should be taken with caution due to the small sample size and the low statistical power of the tests.

In terms of limitations, the present study only provides information about 13 patients. Measuring sex hormones on two alternate days throughout a whole menstrual cycle is demanding on the subject and expensive, and it is for such reasons that our study was limited to an initial number of 15 subjects.

**CONCLUSION**

In conclusion, withdrawal bleeding with VMP with this regimen (7 days of 100 mg every 12 h vaginal) does not significantly alter circulating androgen levels compared to baseline and can be used to time blood sampling in these patients.

This protocol optimizes schedules in daily clinical practice and offers an easier diagnostic process and clinical management without changing androgen levels.

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