Endolysin: A therapeutic alternative for antibiotic resistant pathogens causing urinary tract infections

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Basit A, Tahir H. Endolysin: A therapeutic alternative for antibiotic resistant pathogens causing urinary tract infections. Clin Nephrol Res 2021;5(6):1-2.

Urinary Tract Infections (UTIs) caused by pathogenic bacteria is the most frequent clinical condition affecting almost 150 million population each year worldwide, resulting in a huge economic losses due to clinical presentation. The UTIs treated with antibiotics show post treatment complications, which include repeated recurrence. pyelonephritis, renal damage, irreversible loss to kidney tissue during retrograde proliferation leading to severe risk of bacteremia with a major risk of antibiotic resistance due to recurring infections. Bacteriophageencoded endolysin have been oftentimes recognized as a promising antibacterial agent to eradicate antibiotic-resistant bacteria due to their host specific nature and less chances of resistance development in bacterial pathogens. Endolysin kill bacterial

DESCRIPTION

Urinary Tract Infections (UTIs) caused by common bacterial pathogens is the most frequent clinical condition affecting almost 150 million population each year worldwide [1], from a neonate to a geriatric individual [2]. UTIs resulted in 3.5 billion U.S \$ per year economic losses due to clinical presentation of this disease.

Both gram negative and positive bacteria cause UTIs along with certain fungi [3]. Most common causative agent for UTIs is Uropthgenic Escherichia Coli (UPEC), while other pathogenic bacteria include Klebsiella pneumonia, Pseudomonas aeruginosa, Staphylococcus arophyicus and Staphylococcus aureus [4]. The improper and uncontrolled use of antibiotics has resulted in the development of antimicrobial resistance world-wide. Most of the patients with symptomatic UTIs opt for antibiotic treatment, which in the long run alter the normal microbial flora of reproductive tract and gastro intestinal tract hence leading to development of multidrug resistant pathogens. When the normal flora is disrupted by antibiotics, a gap in the niche is generated, which increase the risk of colonization of drug resistant uropathogenic bacteria causing a higher mortality rate [5].

Most importantly, MDR gram negative pathogens like clinical Klebsiella and Pseudomonas isolates has been nominated as major uropathogens in hospital acquired infections, which has raised therapeutic challenges [6, 7]. This increased incidence of drug resistant uropathogens is an emerging concern for medical fraternity due to limitation of treatment options, which are expensive and still does not avoid the risk of death due to systemic infection. In underdeveloped countries where health care system is not advanced and diagnosis is made without the availability of urine test culture or antimicrobial susceptibility, the emergence of MDR is spiking [8,9]. This continues emergence of MDR uropathogenic strains is mostly seen in the countries with low socioeconomic impact including poverty, ignorance, unhygienic practices and substandard drugs availability which worsen the situation in the health care management categorizing it a public health issue [10,11]. pathogens by hydrolyzing peptidoglycan lyer in cell wall. Though large number of lytic enzymes has been reported against various MDR gram positive bacteria, very few reported against gram negative bacterial pathogens, due to their outer membrane, which acts as a barrier to cross by endolysin to hydrolyze peptidoglycan. Therefore, treatment of MDR gram negative pathogens through lytic enzymes is high challenging. However, engineering of endolysin by fusion of membrane rupturing protein can increase its access to the target site in cell wall. The fusion molecule will increase the host range and provide a path to treat common UTIs caused by gram negative and positive pathogens. This commentary is focusing on how, endolysin can be effectively modified to increase their efficiency against Multi drug resistant bacterial pathogens.

Key Words: Urinary tract infections (UTIs); Endolysin; Multi drug resistance (MDR); Protein engineering

The increased microbial susceptibility of uropathogenens to antimicrobials is attributed to the fact that most of these gram negative pathogens falls under the classification of Extended Spectrum-Beta-Lactamases (ESBLs). These ESBLs presenting Gram Negative Bacteria (ESBLs-GNBs) are the primary culprits involved in UTIs leading to inflexibility of empirical treatment options which worsen the prognosis of condition. These ESBLs-GNBs purpose a significant challenge in the pragmatic antibiotic selection by physicians due to their resistance to wide range of antibiotics (aminoglycosides, cephalosporins and quinolones) [12,13]. The growing prevalence of these ESBLs gram negative uropathogens limit the treatment preferences and pose a unanimous public health implication [14]. The antibiotics are struggling to keep up the pace in this growing phase of pathogenic resistance and acquired mutations.

To provide an alternative therapeutic approach for UTIs, bacteriophage derived endolysins can be of high significance in terms of its efficacy and host specificity. The characteristic features of phage derived endolysin such as high binding specificity and a lower chance of developing resistance have provided promise for endolysins as potential antimicrobial agent [15]. To date, many phage endolysins have been successfully applied as antimicrobial agents to treat P. aeuroginosa, Enterococcus faecalis/faecium, Acinetobacter and E. coli. This kind of alternative may reduce the overuse of antibiotics in the future [16]. However, the major challenge in application of endolysin in the treatment of gram negative pathogens is the presence of outer membrane, which does not allow endolysin to reach cell wall, therefore, accessory proteins such holin are required to rupture the cell wall. Holin rupture the outer membrane to create pores, which allow endolysin to lyse peptidoglycan layer in cell wall [17].

In a recent study, Basit et al., expressed an engineered variant of endolysin from RL phage of Pseudomonas aeruginosa (MDR), by fusing holin at it N terminus. The engineered endolysin showed significant antibacterial activity against a wide range of clinically challenging MDR strains such as Pseudomonas aeruginosa, Klebsella pneumonia, E. coli and Methicillin Resistant Staphylococcus Aureus (MRSA), which are mainly involved

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Received date: September 20, 2021; Accepted date: October 4, 2021; Published date: October 11, 2021

This open-access article is distributed under the terms of the Creative Commons Attribution Non-Commercial License (CC BY-NC) (http:// creativecommons.org/licenses/by-nc/4.0/), which permits reuse, distribution and reproduction of the article, provided that the original work is properly cited and the reuse is restricted to noncommercial purposes. For commercial reuse, contact reprints@pulsus.com involved in UTIs. The holin fused endolysin variant was more active than the simple endolysin, indicating that holin provide access to endolysin to hydrolyze bacterial cell wall [18]. Although many other membrane permeablizers have also been reported to increase lytic activity of endolysin against gram negative pathogens, however, their adverse effects on normal flora limit their in *vivo* clinical application. Therefore, fusion of holin with endolysin in different orientations will not only reduce the side effects on normal flora due its host specific nature, but also increase the bacterial killing efficiency. Various protein engineering methods such as directed evolution [19] or de novo protein design approach [20] can be employed to further enhance the bactericidal efficiency, thermal and pH stability of the holin fused endolysin, to assure their in *vivo* application in treatment of multi drug resistant UTIs.

REFERENCES

- Stamm WE, Norrby SR. Urinary tract infections: disease panorama and challenges. The Journal of infectious diseases. 2001; 183 Suppl. 1:S1-4.
- Kunin C. Urinary Tract Infections in Females. Clinical infectious diseases : an official publication of the Infectious Diseases Society of America. 1994. 18(1): 1-10; quiz 11-2.
- 3. Flores-Mireles AL, Walker JN, Caparon M, et al. Urinary tract infections: epidemiology, mechanisms of infection and treatment options. Nature Reviews Microbiology. 2015. 13(5): 269-284.
- Foxman B. Urinary tract infection syndromes: occurrence, recurrence, bacteriology, risk factors, and disease burden. Infectious Disease Clinics. 2014. 28(1): 1-13.
- Laupland K, Ross T, Pitout JDD, et al. Community-onset urinary tract infections: a population-based assessment. Infection. 2007. 35(3): 150-153.
- Alzohairy M Khadri H. Frequency and Antibiotic Susceptibility Pattern of Uro-Pathogens Isolated from Community and Hospital-Acquired Infections in Saudi Arabia-A Prospective Case Study. Journal of Advances in Medicine and Medical Research. 2011: 45-56.
- Sedighi M, Halajzadeh M, Ramazanzadeh R, et al. Molecular detection of β-lactamase and integron genes in clinical strains of Klebsiella pneumoniae by multiplex polymerase chain reaction. Revista da Sociedade Brasileira de Medicina Tropical. 2017. 50: 321-328.
- Siegel JD, Rhinehart E, Jackson M, et al. Management of multidrugresistant organisms in health care settings. 2006. American Journal of Infection Control. 2007. 35(10): S165-S193.
- Lee SY, Park YJ, Yu JK, et al. Prevalence of acquired fosfomycin resistance among extended-spectrum β-lactamase-producing Escherichia coli and Klebsiella pneumoniae clinical isolates in Korea

and IS 26-composite transposon surrounding fosA3. Journal of Antimicrobial Chemotherapy. 2012. 67(12): 2843-2847.

- Qadeer N, Durrani MA, Iqbal S, et al. Multidrug Resistant Escherichia coli and Klebsiella pneumoniae Causing Urinary tract Infection in Pregnant Women. International Journal of Pathology. 2018: 45-49.
- Jalalpour S. Antibiogram pattern in extended spectrum beta lactamase nano enzyme producing gram negative bacilli in Iranian urinary tract infection. African Journal of Pharmacy and Pharmacology. 2012. 6(12): 899-903.
- Osthoff M, McGuinness SL, Wagen AZ, et al. Urinary tract infections due to extended-spectrum beta-lactamase-producing Gram-negative bacteria: identification of risk factors and outcome predictors in an Australian tertiary referral hospital. International Journal of Infectious Diseases. 2015. 34: 79-83.
- Doi Y, Arakawa Y. 16S ribosomal RNA methylation: emerging resistance mechanism against aminoglycosides. Clinical Infectious Diseases. 2007. 45(1): 88-94.
- Naber K, Llorens L Kaniga K, et al. Intravenous doripenem at 500 milligrams versus levofloxacin at 250 milligrams, with an option to switch to oral therapy, for treatment of complicated lower urinary tract infection and pyelonephritis. Antimicrobial Agents and Chemotherapy. 2009. 53(9): 3782-3792.
- Briers Y, Lavigne R. Breaking barriers: expansion of the use of endolysins as novel antibacterials against Gram-negative bacteria. Future Microbiology. 2015. 10(3): 377-390.
- Carvalho C, Costa AR, Silva F, et al. Bacteriophages and their derivatives for the treatment and control of food-producing animal infections. Critical Reviews in Microbiology. 2017. 43(5): 583-601.
- 17. Tamai E, Yoshida H, Sekiya H, et al. X-ray structure of a novel endolysin encoded by episomal phage phiSM 101 of C lostridium perfringens. Molecular Microbiology. 2014. 92(2): 326-337.
- Basit A, Qadir S, Qureshi S, et al. Cloning and expression analysis of fused holin-endolysin from RL bacteriophage; Exhibits broad activity against multi drug resistant pathogens. Enzyme and Microbial Technology. 2021. 149: 109846.
- Basit A, Tajwar R, Sadaf S. et al. Improvement in activity of cellulase Cell2A of Thermotoga neapolitana by error prone PCR. Journal of Biotechnology. 2019. 306: 118-124.
- 20. Basit A, Karim AM, Asif M, et al. Designing Short Peptides to Block the Interaction of SARS-CoV-2 and Human ACE2 for COVID-19 Therapeutics. Frontiers in Pharmacology. 2021. 12: 2310.