

# Generation of blaNDM-1 super MDR gene by multiple rearrangement of other metallo-beta-lactamases like GIM-1, IMP-1 and VIM-2 including many IS-elements

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## ABSTRACT

Discovery of blaNDM-1  $\beta$ -lactamase in plasmids of *Escherichia coli* and *Pseudomonas aeruginosa* and unresponsiveness of such infections to most antibiotics including imipenem and doripenem shocked the whole drug industry in 2009 as well as discovery of mcr-1 gene in 2016 orchestrated such horror being colistin drug was useless now. We have checked such horror in Ganga River water which contains superbugs, resistant to most common antibiotics including methicillin, lomefloxacin, amikacin, imipenem, linezolid and vancomycin. Beta-lactamases (TEM, OXA, CTX-M, CMY, GIM, VIM, NDM, KPC), drug modifying acetyl/phospho/adenyl transferases and drugs efflux proteins (tetA/C, mexAB/CD/EF) are mostly diversified and mutated creating hundred to thousand isomers. Many MDR

genes are accumulated now in large MDR conjugative plasmids and it is almost impossible to PCR identify all MDR genes in any clinical isolate for better prognosis and therapeutics. From *in silico* protein alignment studies, we concluded that blaNDM-1 gene might be originated from the rearrangement of other Metallo- $\beta$ -lactamases genes present in same plasmids or multiple integrons. This conclusion supports our previous studies with Kolkata superbugs *Escherichia coli* KT-1\_ MDR and *Pseudomonas aeruginosa* DB-2\_ MDR which contain multiple small as well as large plasmids with many diverged MDR genes and transposons. Presence of ThiX gene (in pKOX\_R1 plasmid) involved in Thiamine biosynthesis suggests the bacteria need vitamin synthesizing enzymes in plasmid if we continue high dose complex antibiotic intake. Thus many IS-elements and transposons are assembled in MDR plasmids for new MDR gene creation to save symbiotic relation between gut microbiota and human metabolosome.

**Key Words:** NDM-1  $\beta$ -lactamase; MDR gene evolution; Metallo- $\beta$ -lactamases; blaGIM-1; blaIMP-1; blaVIM-1; Gene rearrangement

## INTRODUCTION

$\beta$ -lactamases have been grouped into four major classes (A–D) based on sequence homology (1) but diverged more profoundly as described recently (2,4). Classes A, C, and D use an active site serine as a nucleophile. However, the class B metallo- $\beta$ -lactamase (MBL) enzymes use bound zinc atoms at the active site (5). Millions of MDR genes have been sequenced that includes mostly bla genes (TEM, SHV, CTX-M-1/2/9, OXA2/23/48/58, VIM, CMY, GIM, VIM etc.), drug modifying genes like catB3, aadA1, aacII-a, aph-Ib and diverse range of drug transporter genes like acrAB, macA/B, mexCD/EF, bcr, bmr, tetA/B, mcr, norA and mtrCDE (6) as well as arr3, sul1/2 and mcr-1. Drug industry is always run to discover new antibiotic derivatives to overcome the actions of MDR genes located in plasmids as well as chromosome. As for example, cefotaxime is good in case of blaTEM containing plasmids but blaCTX-M enzyme lyses cefotaxime and new imipenem drug is introduced. Soon, blaVIM or blaIMP enzymes are developed that destroy imipenem and recently (2009) discovered blaNDM1 could destroy all penicillins, cephalosporins and carbapenems posing a threat to drug industry when no one want to invest in new drug discovery. Lastly, we introduced  $\beta$ -lactamase inhibitor (cavulinate, sulbactam, avibactam) combination therapy but problems continued. Most devastating fact, carbapenems are clustered in large conjugative plasmids in presence of ESBL enzymes like KPC-2, VIM, OXA-23, OXA-210 and diverged drug acetyl/phospho transferases including PBPs and many drug efflux genes (6). Those bacteria are named as superbugs and such infections must be treated in the developed countries like UK and USA (7-9). We are now studying the mutational profiles of bla genes in the isolated MDR-bacteria from Ganga River and Rain water of Kolkata [in press].

*K. pneumoniae* 151kb plasmid (pKP048) (accession no. FJ628167) contains MDR genes like macrolide ABC transporter, floquinolone resistant gene (qnrB4), mercury resistant gene (MerE), sulphonamide resistant protein (sul1) and aph gene for aminoglycoside phosphotransferases (10). blaKPC1 gene was located in *K. pneumoniae* plasmids (accession nos. NC\_022078, NC\_014312, JX283456 and KF954759) and blaNDM1 gene was also located

in many large conjugative plasmids (CP009116, JN420336 and AP012055) (11). Plasmid pKOX\_R1 contains metal resistant genes and many ABC, MFS, AAA drug transporters as well as common MDR genes like cat, sul1, aac3'-IId, FosA3, ANT, aph and NDM-1, CTX-M-3 and SHV-12  $\beta$ -lactamases. Most importantly such plasmids has less Tra genes but 16 types of inserted IS-elements and transposons. We investigated the nature of NDM-1 gene creation here as previously we have detected a homology between VIM-2 and NDM-1 (2-4). We conclude that a critical message has indeed created in bacteria and gut luminal cells to preserve symbiotic relation by continuing synthesis of new MDR genes and acquiring other genetic changes. IS elements, transposases, integrases, recombinases are highly assembled in MDR plasmids and may contribute to the efficient MDR gene creation and AMR void. Interestingly, blaGIM, blaVIM and blaIMP metallo  $\beta$ -lactamases may act as progenitor of super MDR Gene, blaNDM-1.

## MATERIALS AND METHODS

NCBI databases were retrieved using the BLAST programmes ([www.ncbi.nlm.nih.gov/blast](http://www.ncbi.nlm.nih.gov/blast)). The complete genes of blaGIM, blaVIM, blaIMP or blaNDM1 were sequenced in plasmids and were analyzed by Seq-2 programme of BLAST (NDM-1 Protein Ids. AGL09203; AEA41876). Multalin protein sequence software was used to get the nature of conserved sequences among metallo-class B  $\beta$ -lactamases (12). Sometime, diverged sequences are manually cut and paste into align position in MS word so that it is appeared both sequences have similarity. For retrieving any nucleotide like blaGIM-1 or blaVIM, we type the same at the NCBI port ([www.ncbi.nlm.nih.gov/nucleotide](http://www.ncbi.nlm.nih.gov/nucleotide) or Protein) and to BLAST search to type the accession number for protein or DNA into BLAST port ([https://blast.ncbi.nlm.nih.gov/Blast.cgi?PAGE\\_TYPE=BlastSearch](https://blast.ncbi.nlm.nih.gov/Blast.cgi?PAGE_TYPE=BlastSearch)) (13,14).

## RESULT AND DISCUSSION

Multiple align data suggests that many MBL  $\beta$ -lactamases have some interesting similarities to the blaNDM-1 protein that degrades all penicillin, cephalosporin and carbapenem drugs (Figure 1). blaNDM-1 and blaKPC-2

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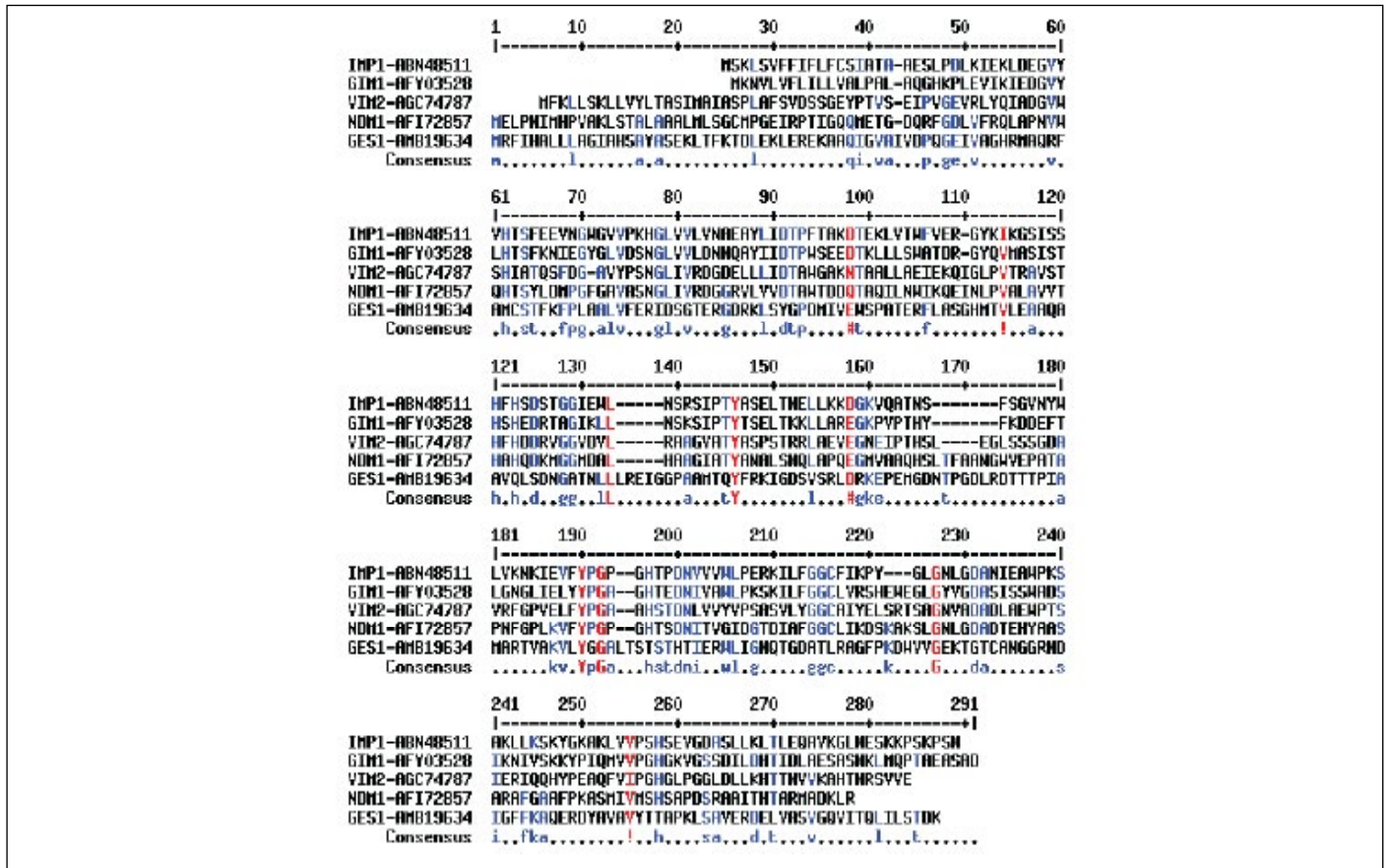


Figure 1) Multi-align of metallo β-lactamases. Different small portions of the alignment shows some similarities: positions 62-93, 121-129, and 195-216. Metallo-beta-lactamases are diverged at the NH2 terminal and no straight similarity zone is observed. We got some hint from such analysis which was confirmed by BLAST seq-2 analysis below

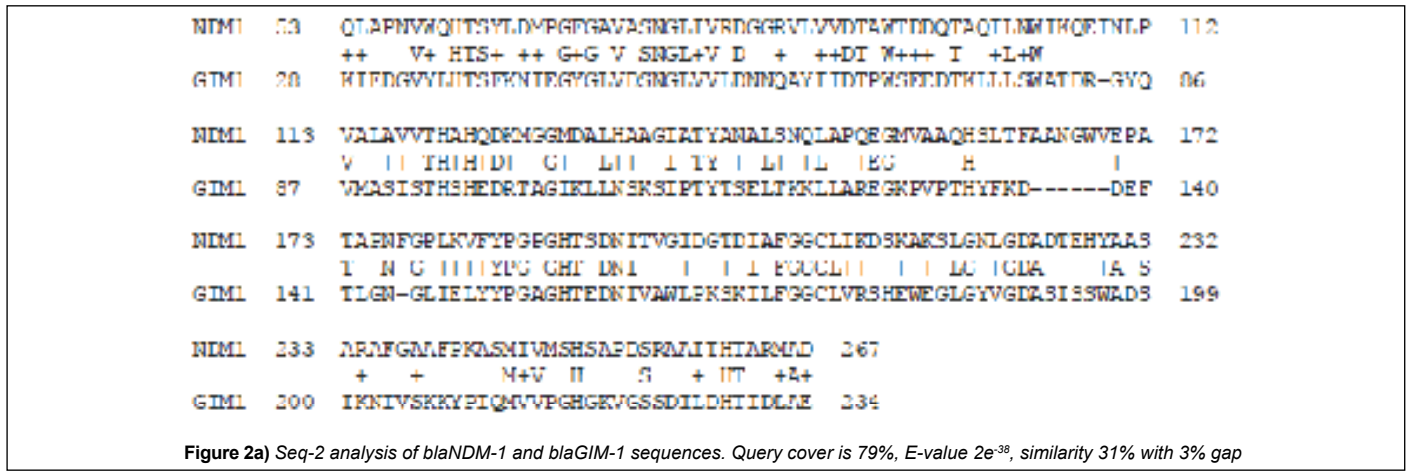


Figure 2a) Seq-2 analysis of blaNDM-1 and blaGIM-1 sequences. Query cover is 79%, E-value 2e<sup>-38</sup>, similarity 31% with 3% gap

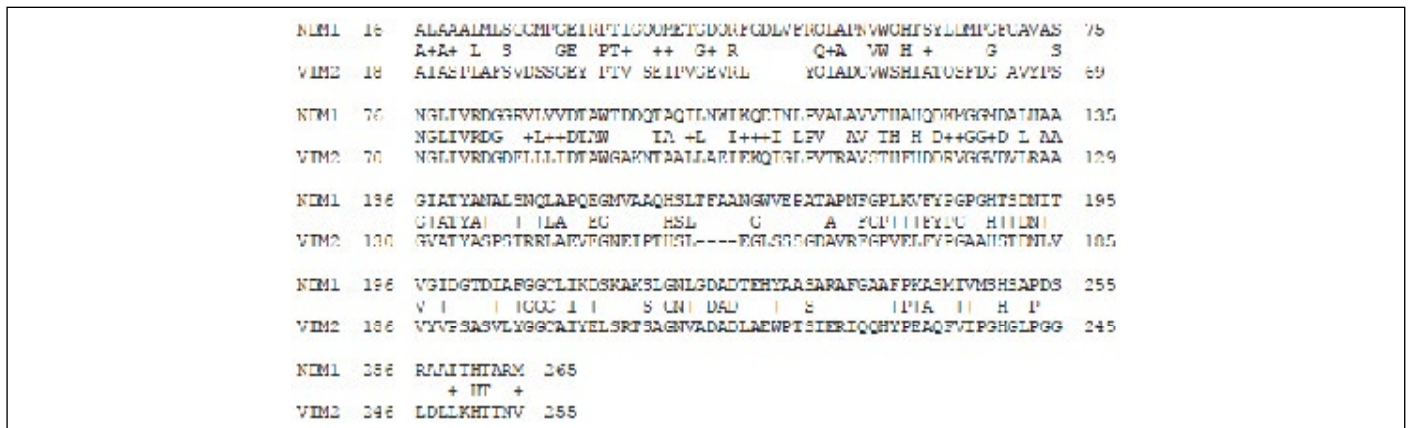


Figure 2b) Seq-2 BLAST analysis of blaNDM-1 and blaVIM-2. Query cover 92%, E-value 2e<sup>-51</sup>, similarity 37% with gap 4%



**Generation of blaNDM-1 Super MDR Gene by Multiple Rearrangement of other Metallo-Beta-Lactamases like GIM-1, IMP-1 and VIM-2 including many IS-Elements**

NDM1	16	ALAPALMISLUMHPEIHTTICQOMHETUDORFQDLVPEOLAINVWQHTSYLLMIPFLAVAS	75
		R+A+ L S GE PT+ ++ G+ R Q+A VW H + G S	
VIM2	18	AIASPLAFVSUSSUEY ITV SEIIPVGEVIL YOLADUVWSHIAIUSPDU AVYIS	69
NDM1	76	NGELIVRDGGRVIVVDITAWTDDQTAQTLNWTKQETNFVVALAVVTHAIQOMHGRGNATHAA	135
		NGELIVRDG +L++DLW IR +L I++I LFW NV TH H D +GG+D L AA	
VIM2	70	NGELIVRDGDFITLITDFAWGAFTAAFLARIEKQTRFVTRAVSTHIFIDRVRGKWTAA	129
NDM1	196	GIATYANALSNQLAPQEGMVARQHSLSLTFANGWVEEXTAPNFGPLKVFYFGPGHSTDNIT	195
		GIATYAI I IIA EG HSL G A FQIITIEYIC HILNI	
VIM2	130	GVATYAGPQTRRLAKVFGNFIPTUST----EGLSGGQDAVRFGPVELFVFAAHTDMLV	165
NDM1	196	VGIDGTDIARGGCLIKDSKAKSLGNLGDADTEHYRASAFAFGAFAFKASKIVMSHAPDS	255
		V I I IGGC I I S UNL DAD I S IPIA I I H P	
VIM2	186	VYVFSASVLYGGCAIYELSRFSAGNVADADLAEWPTSIERIQQHYPEAQFVTPGHSLPGG	245
NDM1	286	RFLIITHIARM	285
		+ IT +	
VIM2	246	LDLLKHTINV	285

**Figure 2c) Seq-2 BLAST analysis of blaNDM-1 and blaIMP-1. Query cover 84%, E-value 3e-43, similarity 34% with 4% gap**

(a)	GIM-1	182	WEGLOVVDASISSWADSIKIVSKYPTONVTC	216	GIM-1	207	KYFIQHWVCHS	218
			WE + GD W SE K R P++ P				KY Q V+RG G	
Tn1696	484	WEHINLIGD---YWRSSMVGAGKFRFLRFLRFA	515	IS3000	72	KVYVQVLRGGG	83	
(b)	VIM-2	50	GWSHIAIUSGDFRWY	67	VIM-2	170	VELFYDGAHSIDNLVWY	187
			G W I S DG +				+E YP A HS V+	
Tn1696	195	GWGGDGTISSDGGNF	210	Inu1	216	LIRKYPRAHSKFWFWF	233	
(c)	NDM-1	143	ALSNQLAPQEGMVARQHSLSLTFANGWVEPATA	160	NDM-1	186	FGPGHSTDNIT	196
			A S A G+V Q+S				FGPG N+T	
IS4321	287	ASSRIFAARTGLVPRDYS	284	IS3000	80	FGPGCREFFLI	90	
(d)	NDM-1	90	DTAWTDDOTAQILNWKICEINLIVVALAV		VTHARQKRMCCMDALHAACIATYANAL		144	
			DTR D+I + + INL +		I+A K+ + A H TY+ AL			
Tn1696	128	DTR--KDKTILLITLIDADAINLGLTYCAESCPGTTVA		---KLSWLOAWHIRD-ETYSIAL			181	
NDM-1	143	SNQLAPQEGMVARQHSLSLTFANGWVEPATA		-----FNFG--FLKV			182	
			+ +V AQ FA N W + I+		P +G P +			
Tn1696	102	AR-----VWPAQFRQPFAGI--WDRGTTGSDRQNFRTGSHAESTYGHINPKVSRSPGRT		233				
NDM-1	183	FYFGPGHSTDNIT		-----ITVGDGTDIARGGCLIKDSKAKSLGNLGDADTEHYRASA			233	
			FY H SD + VGI + G L +S +		KHY +A			
Tn1696	281	FYT HISDQYAFSAKVVWVGLRDSIYVLDGLLYHESLRL		-----EKHYTDLA			281	

**Figure 3) Demonstration of some homology between IS-elements and metallo-β-lactamases. Comparative analyses were done between IS-elements (IS4321, Tn1696, TnpA, Int1, IS3000 and ISEcp1) present in NDM-1 plasmid (pNDM-US, accession no. CP006661) and GIM-1, NDM-1 and VIM-2 β-lactamases. Analysis suggested such transposons may help to form blaNDM-1 MDR gene induced by antibiotics involving gut microbiota**

(a)	NDM-1	90	DTAWTDDOTAQILNWKICEINLIVVALAV		VTHARQKRMCCMDALHAACIATYANAL		144
			DTR D+I + + INL +		I+A K+ + A H TY+ AL		
Inu	600	DTR--KDKTILLITLIDADAINLGLTYCAESCPGTTVA		---KLSWLOAWHIRD-ETYSIAL			600
NDM-1	143	SNQLAPQEGMVARQHSLSLTFANGWVEPATA		-----FNFG--FLKV			182
			+ +V AQ FA N W + I+		P +G P +		
Tn3	657	AR-----VWPAQFRQPFAGI--WDRGTTGSDRQNFRTGSHAESTYGHINPKVSRSPGRT		708			
NDM-1	183	FYFGPGHSTDNIT		-----ITVGDGTDIARGGCLIKDSKAKSLGNLGDADTEHYRASA			233
			FY H SD + VGI + G L +S +		KHY +A		
Tn3	709	FYT HISDQYAFSAKVVWVGLRDSIYVLDGLLYHESLRL		-----EKHYTDLA			786
(b)	NDM-1	83	GGEVLVVDRAWTDDQTAQILNWKICE		108		
			GU V V+ + QIL +KQ+				
IS-91	454	GGEVETVASTEDPKVIFQITLKHNRQK		479			
(c)	NDM-1	150	PODGMVARQHSLSLTFANG---WVEPATA		----APNDG		170
			P AAI L A G W IKT AITIC				
IS 6	107	PTRSAKAAKRELCALRCLRHWEKDATINTDKA2SYC		143			

**Figure 4) Seq-2 BLAST align of IS-elements located in NDM-1 plasmid (pKOX\_R1; accession no. NC\_018107) with blaNDM-1. Klebsiella michiganensis MDR plasmid pKOX\_R1 has >150 genes and 23 IS-elements insertions (16 types). Only Tn3, IS-6, IS-91 show some homology than others (IS-4, IS-5, IS-9, IS-110, IS-630, IS-1182, ISKox1, ISKpn21 etc. Notably pKOX\_R1 plasmid contains drug transporter (accession no. WP\_001549953), arsenic, mercury, tellurium resistant determinants and many DUF domain proteins. Many MDR genes like cat, aac3-III, ABC transporter, mph-E, sul1, dhfr, CTX-M-3, aacA4, fosA3, SHV-12 including 16S rRNA G<sup>405</sup> N<sup>7</sup>-methyl transferase (ArmA gene) are scattered in the plasmid backbone**

containing plasmids are emerging in India and such superbugs are very deadly as unresponsive to most antibiotics (15-17). We used seq-2 BLAST (www.ncbi.nlm.nih.gov/blast /nucore/protein) analysis to pinpoint the homology between GIM-1, VIM-2 and IMP-1 β-lactamases. NDM-1 at position '74' and VIM-2 at position '68' have "SNGLIVRDG" nine amino acid similarities and at 110 position within 31 amino acids 20 are identical (65%) suggesting blaNDM-1 gene has partially originated from blaVIM-2 gene (Figure 1; Figure 2a). Where as at 62 aa position of NDM1, we see a conserved motifs (HTSYLDMPGFGAVASNGLIRD) with GIM1 protein sequence with 12aa identity (Figure 2b). The seq-2 align suggests that "VFYPGPGHT" nine amino acids and "LGNLGD" seven amino acids

perfect matches with IMP-1 β-lactamase at 182/218 positions respectively (Figure 2c). and is very interesting as many plasmids carry multiple such genes and may facilitate the generation of blaNDM-1 super protein that destroys all penicillins, cephalosporins and carbapenems (4). In history of hundreds of MDR genes, NDM-1 containing superbugs have created the most horror in society other than drug modifying (cat, aac, aph) and drug efflux genes (mex and tet). In truth, blaNDM1 and KPC2 containing superbugs are increasing in the environment of India (2,9) and many NDM1 mutants have now sequenced (4). When we aligned with KPC2 protein sequence and only "156-AAQHSLSLTFANG" was aligned considerably (125-AVQYSDNAAAN) suggesting KPC-2 was not involved in that case (18).

We further investigated the occurrence of transposons and IS-elements in large as well as small MDR plasmids. Many medium sized plasmids (25-40kb) do contains 4-8 MDR genes and few IS-elements containing transposases, resolvases and integrases (3). We detected that Tn1696, Int1 and IS3000 have some similarity zones to blaNDM-1 and also to blaGIM-1 (Figure 3). In pKOX\_R1 plasmid we found 23 IS insertions and such astonishing result we explored further. Seq2 BLAST analysis detected few stretch of 30-50% homology with blaNDM-1 protein sequence with Tn3, IS-6 and IS-91 sequences (Figure 4). Small plasmids and integrons (see, accession nos. EF375699; KT984195; JX566715; GQ466184; EU597467; DQ310703; NC\_019081) carry VIM-1, GES-1, GIM-1, IMP-1 carbapenemases including ESBL OXA  $\beta$ -lactamases including many integrases and recombinases. Thus, large plasmid (accession nos. AP012055, NC\_018107, KT185451) may generate later and blaNDM-1 gene may be generated in small plasmid which directly combined with conjugative plasmid (10). As for example, isolated imipenem resistant microbes from Ganga River water seem lacking tetracycline, streptomycin and chloramphenicol resistant determinants (unpublished). In truth, imipenem resistant superbugs are rare in the environment (0.002%), although in clinical isolates 5-10% imipenem resistant bacteria are detected. *Klebsiella pneumoniae* large plasmid pCR14\_2 (accession no. NZ\_CP015394) also have inserted many transposons (IS110, IS91, Tn3, IS5, Int1) including clustered MDR genes OXA-2, sul1, CTX-M2 and AAC(3)-IIa.

### CONCLUSION

MDR horror has reached every home of this Earth and WHO advised new interventions to stop this medical catastrophe (11, 19-22). We found that super conjugative plasmids have many MDR genes in single plasmid and also many small plasmids like integrons and IS-elements (23,24). We postulated that a critical message has generated between bacteria and human at the intestine to preserve symbiotic relation in presence of high dose of antibiotics that all are taking since 1940s (25). One high dose of antibiotic kills gut microbiota that synthesize 20 vitamins and complex bio-molecules which are absolutely needed for normal human metabolosome evolving >30000 enzymatic bio-conversions. MDR plasmids contain many transposases, resolvases, topoisomerases and integrases and such enzymes are constantly rearranging genes in bacterial cytoplasm (2, 12). Thus bacteria can make a new MDR gene against new antibiotic in the intestine activated by antibiotic itself and probably within few weeks to save vitamin biosynthesis for human. We proved here that deadly NDM-1 gene is created from plasmids containing blaGIM-1, blaVIM-2 and blaIMP-1 like genes which present in combination with many other genes and IS-elements. We found ThiF gene (in pKOX\_R1 plasmid) that involved in thiamine vitamin biosynthesis indicating MDR bacteria will be acquired all vitamin synthesizing enzymes in MDR plasmids gradually if we will be continued complex antibiotic intake. Such large plasmid has few Tra conjugative genes (conjugative deficient) but contains 23 insertion elements suggesting a great site for MDR gene creation. We believe interventions targeting DNA rearrangement in superbugs may be a alternative target to control superbug spread (26-30).

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