Pro apoptotic effect of neosedumoside III on human hepatocellular carcinoma hepG2 cells and SMMC 7721 cells through G0/G1 phase cell cycle arrest and regulate caspase-cascade pathway

Qing-Qing Zhao^{*}, Xin-Yu Li, Xin-Zhou, Yu-Liu, Feng Qiu

Qing-Qing Z. Pro apoptotic effect of neosedumoside III on human hepatocellular carcinoma hepG2 cells and SMMC 7721 cells th rough G_0/G_1 phase cell cycle arrest and regulate caspase-cascade pathway. J Histol Histopathol Res 2022; 6(3):1-13.

ABSTRACT

Purpose: NeosedumosideIII (Neo) is a megastigmanes and belongs to monocyclic sesquiterpenoids compound with antioxidant, anti-

inflammatory and pharmacological activities. In order to explore the anticancer effect and possible mechanism of Neo, the study examined the antiproliferation and apoptosis effect of Neo against human hepatocellular carcinoma HepG2 cells and SMMC-772 cells and related mechanism *in vitro*.

Methods: The anti-proliferation effect of Neo was detected on HepG2 cells and SMMC-772 cells by MTT assay and IC50 with increasing dose and

INTRODUCTION

Liver cancer is one of the most common tumors in the world and mainly caused by chronic hepatitis, fatty liver and other liver diseases [1] it's the second leading cause of cancer-associated high morbidity and mortality all over the worldwide [2,3]. At present, the proportion of mortality and morbidity of liver cancer showed tendency to ascend and the relative survival rate was reduced that indicated a poor prognosis [4]. Liver cancer is not very sensitive to chemotherapy and radiotherapy, and the mainly used treatment methods are surgical resection. however, liver cancer due to its high invasive nature and most of the patients have developed to advanced stage at the time of diagnosis, lost the best time for treatment [5]. Therefore, the most important task is screening potential and effective compounds for the treatment of liver cancer.

Natural products isolated from Traditional Chinese Medicines are one of the important sources for anti-cancer drug which used as effective cancer treatment in China. NeosedumosideIII (Neo) is a megastigmanes and belongs to a class of monocyclic sesquiterpenoids with antioxidant, antiinflammatory and other pharmacological activities [6,7]. A large number of literature reports monocyclic sesquiterpenoids are widely distributed in nature, and have inhibitory activity for human breast cancer cells MDA-MB-231, human colon cancer cells HCT116, Human liver cancer cells HepG2 and SMMC-7721, etc. [8-10]. Meanwhile, a megastigmanes has inhibitory activity for human lung adenocarcinoma cells A549, human prostate cancer cells PC3 and human breast cancer cells MCF7 [11]. However, Neo as a megastigmanes compound and the inhibition of Human liver cancer cells HepG2 and SMMC-7721 has not been reported, and the protective mechanism of cos against liver cancer is still unclear. Therefore, in order to explore the anti-cancer effect of Neo and its possible mechanism. The study evaluated the effect of Neo against liver cancer and the

time. Cell cycle and apoptosis were detected by flow cytometer. The changes of Bcl-2, Bax, Caspase-3, Caspase-8 and Caspase-9 proteins were detected by western blotting.

Results: The results indicated that Neo could inhibited proliferation of HepG2 cells and SMMC-772 cells *in vitro* and promoted apoptosis, it significantly induced apoptosis of HepG2 cells and SMMC-772 cells arrested cell cycle at G 0/G1 phase in a dose-dependent manner, reduce the expression of Bcl-2 protein, and increase the expression of Bax and Caspase-3, Caspase-8 and Caspase-9 proteins.

Conclusion: Neo could inhibit proliferation and induce apoptosis of HepG2 cells and SMMC-7721 cells *in vivo* which suggested that it might be served as a promising candidate for the treatment of liver cancer.

Keywords: Neosedumoside III; Hepatocellular carcinoma; HepG2 cells and SMMC-772 cells; Mechanism

mechanism of action of Neo on human liver cancer ${\rm HepG2}$ cells and SMMC-772 cells in vitro.

In this study, the growth inhibit effect of Neo was detected on HepG2 cells and

SMMC-7721 cells by MTT assay. Cell cycle and apoptosis analysis were detected by

flow cytometry. The expression of Bax, Bcl-2 and Caspase-3, Caspase-8, and Caspase-9 proteins were analyzed by western blotting, the study were used to analyze whether Neo could be used as a promising candidate drug for the treatment of liver cancer. We found that Neo effectively inhibited cell proliferation and induced apoptosis in HepG2 cells and SMMC-7721 cells and its potential mechanisms underlying the effects, it indicate that Neo might be used as a potential drug for the liver cancer therapy.

MATERIALS AND METHODS

Reagents

Neo was bought from Herbpurify Company (Chendu, China, purity >98%) and dissolved in DMSO and stored at 4°C. Human Hepatocytes L-O2 cells, Human Hepatocellular Carcinoma HepG2 cells and SMMC-7721 cells were purchased from Shanghai Institute of cell biology for Chinese Academy of Sciences (Shanghai, China). Dulbecco's minimum essential medium (DMEM, Gibco), fetal bovine serum (FBS, Gibco), streptomycin and penicillin (Invitrogen) were bought from Wolcavi Company (Beijing, China). Methyl Thiazolylte Trazdium assay kit (MTT), Dimethyl sulfoxide (DMSO), polyclonal antibodies (β-actin, Bax, Bcl-2, Caspase-3, Caspase-8, and Caspase-9) and secondary antibodies (goat-anti mouse), BCA protein kit, Trypsin, RIPA lysis buffer, cell cycle analysis kit and cell apoptosis detection kit were bought from Beyotime Company (Shanghai, China).

Department of Pharmacy, Chongqing Medical University, China

Correspondence: Qing Qing Zhao, Department of Pharmacy, Chongqing Medical University, China, Tel: +8613983814114; Email: qingqin2350@163.com

Received: December 10, 2021, Manuscript No. PULHHR-21-4033; Editor assigned: December 13, 2021, PreQC No. PULHHR-21-4033 (PQ); Reviewed: December 27, 2021, QC No. PULHHR-21-4033; Revised: February 10, 2022, Manuscript No. PULHHR-21-4033 (R); Published: March 01, 2022, DOI: 10.37532/ pulhhr.22.6(3).68-72

 This open-access article is distributed under the terms of the Creative Commons Attribution Non-Commercial License (CC BY-NC) (http:// creativecommons.org/licenses/by-nc/4.0/), which permits reuse, distribution and reproduction of the article, provided that the original work is properly cited and the reuse is restricted to noncommercial purposes. For commercial reuse, contact reprints@pulsus.com

J Histol Histopathol Res Vol 6 No 3 2022

Zhao QQ et al.

Western Bright ECL prime Western-blotting detection reagent was bought from Ding-guo Company (Beijing, China).

Cell culture

Human Hepatocytes L-O2 cells, Human Hepatocellular Carcinoma HepG2 cells and SMMC-7721 cells were grown in DMEM (Gibco) containing 10% FBS (Gibco), and supplemented with streptomycin and penicillin (Invitrogen). Both types of cells were conducted at 37°C in a humidified 5% CO₂ incubator. Subsequently, the control group was cultured with DMSO whereas hepatocellular carcinoma HepG2 cells and SMMC-7721 cells were treated by 2, 4, 8, 16, and 32 µmol/L NEO for 0, 3, 6, 12, 24 and 48 h. For Neo treatment, the final concentration of DMSO added to the cells was <0.1%. In addition 0.1% DMSO group was used as a control.

Cell viability assay

MTT assay were used to assess viability and applied to measure L-O2 cells, HepG2 cells and SMMC-7721 cells viability. Three kinds of cells were plated 5 × 104 cells/well in 96-well culture plates and treated with serial concentrations of Neo (2, 4, 8, 16, 32 μ mol/L) for 48 h (various time periods) and MTT solution (5 mg/mL) was added to each well, DMSO was used as a control. Subsequently, the cell viability is measured by MTT assay; the IC 50 (The half maximal inhibitory concentration) values were calculated by linear-regression analysis and averages of three independent repeats being calculated.

Cell cycle analysis

After HepG2 cells and SMMC-7721 cells were treated with serial concentrations of Neo (2, 4, 8, 16, 32 μ mol/L) and DMSO for 48 h, both types of cells were collected and centrifuged at 1000 g for 5 min to remove the culture medium, the cells were washed twice with PBS for cell cycle analysis. The cells were fixed with 70% ethanol at 4°C for 2 h, and the fixed solution was discarded, then cells resuspended 5 min by PBS and centrifuged at 1000 g for 5 min, PBS was discarded and stained with Propidium Iodide (PI), the cells were darkened at 4 °C for 30 min, and cell cycle was detected by flow cytometry.

Cell apoptosis assay

After HepG2 cells and SMMC-7721 cells were treated with serial concentrations of Neo (2, 4, 8, 16, 32 μ mol/L) and DMSO for 48 h and then cells were washed twice by PBS. Cells were resuspended with 500 μ L binding buffer, Annexin V-FITC (5 μ l) and PI solution (5 μ l) were added and incubated at room temperature for 15 min in the dark, and the cell apoptosis were analyzed by flow cytometry (ACEA Novo-Cyte, Agilent Technologies Inc., USA). According to the instructions of apoptosis kit, the number of cells was calculated by cell analysis software (BD Inc., USA) for data analysis and the experiments were performed in triplicate.

Western blot assay

Human Hepatocellular Carcinoma HepG2 cells and SMMC-7721 cells were analyzed by western blotting. The total protein fractions were isolation and DMEM medium were removed. Subsequently, cells were washed twice by PBS and then lysed using cell by RIPA lysis buffer. The lysates were collected and centrifuged at 12000 g for 5 min at 4°C, protein concentrations were determined using the BCA protein assay. Samples were separated by 10% SDS-PAGE (sodium dodecyl sulfate polyacrylamide gel electrophoresis) and then transferred to PVDF membranes, and blots were blocked with 10% skim milk incubated with the primary antibodies Bax, Bcl-2, Caspase-3, Caspase-8, Caspase-9, anti β -actin overnight at 4°C. Then the membranes were incubated with secondary antibodies conjugated to HRP (Horse Radish Peroxidase), and gray scale analysis used by Image J software platform, the relative protein expression were expressed by the ratio of gray value of Bax, Bcl-2, Caspase-3, Caspase-8, and Caspase-9 protein to β -actin.

Statistical analysis

Data were expressed as the mean \pm SD. Statistical comparisons of results were made using analysis of variance (ANOVA). Significant differences (*P<0.05, **P<0.01) whereas T-tests were used by SPSS software (version 19.0). P values with P<0.05 was the threshold for a statistical difference, whereas P<0.01 was the threshold for a significant difference. The graphs were generated with Origin version 9.0.

RESULTS

Effect of Neo on the proliferation of HepG2 cells and SMMC-7721 cells *in Vitro*

The chemical structure of Neo was shown in Figure 1A, which is a megastigmanes and belongs to monocyclic sesquiterpenoids compound. As shown in Figure 1B, the result of inhibitory effects showed that Neo effectively inhibited HepG2 cells proliferation in a dose-dependent and time-dependent (0, 3, 6, 12, 24, 36 and 48 h) compared with the DMSO group, and the IC50 was 16.15 ± 1.27 μ M. The findings were similar for SMMC-7721 cells (Figure 1C), which also showed that Neo effectively inhibited SMMC-7721 cells proliferation in a dose-dependent and time-dependent (0, 3, 6, 12, 24, 36 and 48 h) compared with the DMSO group, and the IC50 was 15.87 ± 1.31 μ M. In addition, NEO induced relatively lower cytotoxicity in L-O2 cells than in HepG2 and SMMC-7221 cells (Figure 1D) which shows it is safe for normal cells. In a word, other experiment and research were based on these results for further study in vivo.

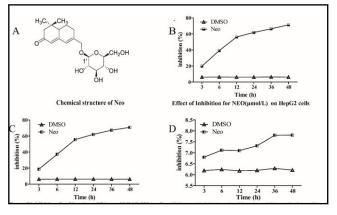
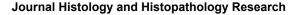


Figure 1: Effect of Inhibition for Neo (µmol/L) on HepG2 cells and SMMC-7721 cells *in Vitro*. A: The chemical structure of Neo. B: Effect of Inhibition for Neo on HepG2 cells. C: Effect of Inhibition for Neo on SMMC-7721 cells. D: Effect of Inhibition for Neo on LO2 cells.

Effect of Neo on cell cycle distribution of HepG2 cells and SMMC-7721 cells *in Vitro*

The control group were cultured with DMSO whereas hepatocellular carcinoma HepG2 cells and SMMC-7721 cells were treated by 2, 4, 8, 16, and 32 μ mol/L NEO for 48 h, the effect of Neo on the cell cycle of hepatocellular carcinoma HepG2 cells and SMMC-7721 cells were detected by PI staining and the results were shown in Figure 2. Compared with the control group, there was no significant change in cell percentage at G2/M stage in Neo group with different concentrations.



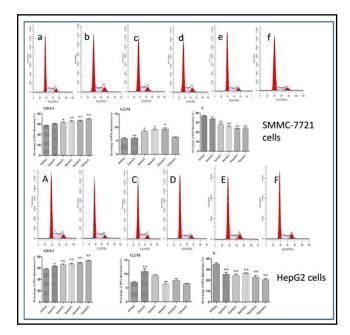


Figure 2: Effect of Neo on cell cycle distribution of HepG2 cells and SMMC-7721 cells in Vitro.

After all group cultured with either DMSO or for Neo 48 h, HepG2 cells and SMMC-7721 cells harvested fixed with ethanol and stained with PI, DNA content was determined by flow cytometery. The percentage of both cells at each phase of cell cycle was analyzed by Origin 9.0. A: DMSO, B: 2 μ mol/L cos, C: 4 μ mol/L cos, D: 8 μ mol/L cos, E: 16 μ mol/L cos, F: 32 μ mol/L cos. The lowercase letters have the same concentration as uppercase letters. Lowercase letters presents SMMC-7721 cells while uppercase letters present HepG2 cells. Data was expressed as mean ± standard deviation from triplicate determinations. *P<0.05, **P<0.01 vs. control.

While cell percentage at G0/G1 stage was significantly increased and cell percentage at S stage was significantly decreased. The change proportion of G0/G1 phase of HepG2 cells were caused by the different concentrations of Neo for (57.93+2.32) %, (63.34+1.22) %, (65.93+2.36) %, (67.02 +2.03) %, (69.25+1.33) % and (72.92+1.43) %. And the change proportion of G0/G1 phase of SMMC-7721 cells were caused by the different concentrations of Neo for (57.26+1.03) %, (60.85.15+1.24) %, (63.60+1.60) %, (65.36+2.56) %, (66.90+1.48) % and (70.05+1.75) %. It was shown that Neo could arrest the cell cycle of hepatocellular carcinoma HepG2 cells and SMMC-7721 cells at G0/G1 phase and has a certain inhibitory effect on cell migration to G2/M stage.

Effect of different dose Neo on apoptosis in HepG2 cells and SMMC-7721 cells

To investigate the Neo induced apoptosis of hepatocellular carcinoma HepG2 cells and SMMC-7721 cells, the control group were cultured with DMSO whereas hepatocellular carcinoma HepG2 cells and SMMC-7721 cells were treated by 2, 4, 8, 16, and 32 µmol/L NEO for 48 h, and the results were shown in Figure 3 and Figure 4. The apoptosis rate of HepG2 cells were caused by the different concentrations of Neo for (2.28 \pm 0.68) %, (6.86 \pm 1.30) %, (13.01 \pm 3.23) %, (20.08 \pm 3.10) %, (31.97 \pm 2.88) % and (50.67 \pm 5.89) %. And the apoptosis rate of SMMC-7721 cells was caused by the different concentrations of Neo for (1.82 \pm 0.30) %, (4.82 \pm 1.89) %,(11.44 \pm 2.54) %, (15.62 \pm 2.38) %, (30.55 \pm 2.92) % and (43.82 \pm 3.11) %. It was shown that the apoptosis rate of hepatocellular carcinoma HepG2 cells and SMMC-7721 cells increased with the increase of Neo concentration, which also indicated the potential anticancer mechanism of Neo.

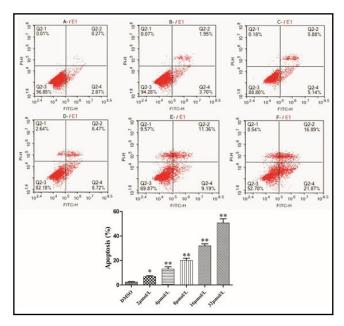


Figure 3: Effect of Neo on apoptosis in HepG2 cells in Vitro.

After all group cultured with either DMSO or for Neo 48 h, the HepG2 cells were stained with FITC-conjugated Annexin V in a buffer containing PI and analyzed by flow cytometry. The percentage of HepG2 cells categorized as apoptotic were analyzed by SPSS 19.0. A: DMSO, B: 2 μ mol/L cos, C: 4 μ mol/L cos, D: 8 μ mol/L cos, E: 16 μ mol/L cos, F: 32 μ mol/L cos. Data was expressed as mean ± standard deviation from triplicate determinations. *P<0.05, **P<0.01 vs. control.

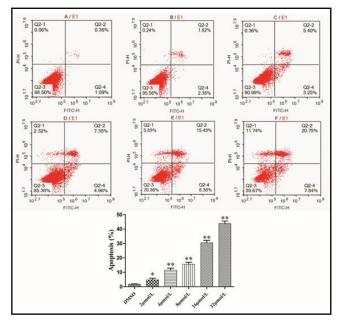


Figure 4: Effect of Neo on apoptosis in SMMC-7721 cells in Vitro

Effect of the Neo on the expression of Caspase-3, Caspase-8, and Caspase-9, Bax, and Bcl-2 in HepG2 cells and SMMC-7721 cells in Vitro.

As shown in Figure 5 and Figure 6. The expression of Bcl-2 family proteins and the caspase family proteins have the same change trend in both HepG2 cells and SmMC-7721 cells with the adding of Neo. It was found that Neo induced apoptosis of HepG2 cells and SmMC-7721 cells by up-regulating Bax, Caspase-3, Caspase-8 and Caspase-9 proteins and down-regulating Bcl-2 protein expression. All these changes were dependent upon the dose of Neo.

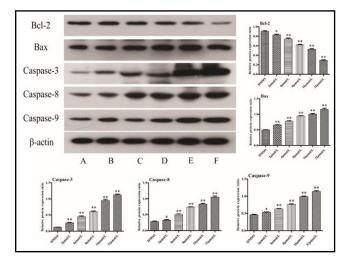


Figure 5: Effect of the Neo on the expression of Caspase-3, Caspase-8, and Caspase-9, Bax, and Bcl-2 in HepG2 cells cells in Vitro.

Western blotting analysis was used to detect protein expression levels of apoptotic markers on HepG2 cells, β -actin served as an internal control. A: DMSO, B: 2 µmol/L cos, C: 4 µmol/L cos, D: 8 µmol/L cos, E: 16 µmol/L cos, F: 32 µmol/L cos. Data was expressed as mean ± standard deviation from triplicate determinations. *P<0.05, ***P<0.01 vs. control.

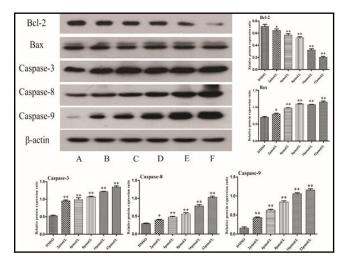


Figure 6: Effect of the Neo on the expression of Caspase-3, Caspase-8, and Caspase-9, Bax, and Bcl-2 in SMMC-7721 cells *in Vitro*.

Western blotting analysis was used to detect protein expression levels of apoptotic markers on SMMC-7721 cells, β actin served as an internal control. A: DMSO, B: 2 µmol/L cos, C: 4 µmol/L cos, D: 8 µmol/L cos, E: 16 µmol/L cos, F: 32 µmol/L cos. Data was expressed as mean ± standard deviation from triplicate determinations.

DISCUSSION

The sesquiterpenes are widely distributed in nature. The monocyclic sesquiterpenoids are abundant in sesquiterpenes with many physiological activities including anti-inflammatory, antioxidant, anti-ulcer, cytotoxic activities and other physiological activities [12-14], among which anti-inflammatory and cytotoxic activities are the most prominent. There are many studies have shown that megastigmanes in different cancer cells have different cytotoxic activities [10] and Neo is a type of megastigmanes compound, in order to further verify the antitumor activity of Neo, the cytotoxic activities of Neo against HepG2 cells and SMMC-7721 cells in *vitro* was investigated. In this study, it was showed that Neo could inhibit the proliferation of HepG2 and SMMC-7721 cells and in a dose-dependent.

4

Carcinogenesis is a long and multistep process affected by many factors. The cell cycle is a primary factor which is a very sophisticated process and different compounds have different effects on the cell cycle, the G0/G1 and G2/M phases are critical points in this process. It was found that the volatile oil significantly inhibited the proliferation of hepatocellular carcinoma HepG2 cells and SMMC-7721 cells cultured in vitro may be due to the increase of G2/M phase and S phase, accompanied by the decrease of G 0/G1 phase [15]. The effect of Rosiglitazone may be prevent the retention of hepatocellular carcinoma HepG2 cells in G_0/G_1 phase and inhibit the cell migration to G2/M phase to a certain extent [16]. The Drugcontaining serum of Clerodendrum Bunge blocks hepatocellular carcinoma MHCC97-H cells from G2/M phase to G0/G1 phase and further reducing S phase, thereby interfering with DNA replication to inhibit cell proliferation [17]. Therefore, it is important to analyze the effect of substances on cell proliferation by detection for the cell cycle, our study found that the Neo could block the cell cycle arrest in G0/G1 phase, it indicated that Neo can block the cell cycle of both types of cells to some extent and it was a great significance for the study of anticancer mechanism.

Apoptosis is a physiological process of cells. In recent years, many studies have shown that the occurrence and development of tumors were related and may be caused by abnormalities in the process of cell apoptosis [18,19]. Bcl-2 proteins and Bax proteins belongs to Bcl-2 family and then were key factors in the process of cell apoptosis which were expressed abnormally in various tumors, and then have become one of the targets of anticancer drugs [20]. Cell apoptosis may be induced by inhibition of Bcl-2 or activation of Bax [21], the lower the ratio of Bcl-2/Bax and the higher mortality of the cell, therefore, the up-regulation of Bax protein expression or the down-regulation of Bcl-2 protein expression might be related to the apoptosis of various tumor cells including liver cancer [22]. In addition, the study indicated that there are two apoptotic pathways including the "intrinsic" cytochrome C/Caspase-9 pathway and the "extrinsic" Caspase-8 pathway [23], caspase family proteins were also play a key role of cell apoptosis. Caspase-9 as a promoter of caspase which were necessary for apoptosis signal through mitochondrial pathways and activated on apoptotic complex, and thus activate caspase-3 [24]. Caspase-8 was also as a promoter of caspase and thus activate caspase-3 eventually leads to cell apoptosis [25]. When Caspase-3 was activated by upstream pathway and as a key "killer" of cell apoptosis [26] and induces cascade reactions leading to cells apoptosis [27]. Research suggests that costunolide can activate caspase-3 to mediate apoptosis of esophageal cancer cells by up-regulation of Bax protein expression and down-regulation of Bcl-2 protein expression [28], breast cancer McF-7 cells apoptosis induced by curcumol through regulating the expression of Bcl-2 protein and Bax protein and down-regulating the ratio of Bcl-2/Bax [29], and many other studies have shown that inducing apoptosis was important for inhibiting the development of liver cancer [30]. Therefore, the study investigated the effect of Neo on the expression levels of the key regulators of apoptotic pathways in HepG2 cells and SMMC-7721 cells including Bax, Bcl-2, Caspase-3, Caspase-8 and Caspase-9. In this study, western blotting analysis showed that Neo induced apoptosis of HepG2 cells and SMMC-7721 cells by up-regulating the expression of Bax, Caspase-3, Caspase-8 and Caspase-9 proteins, and downregulating the expression of Bcl-2 protein. The results of the study were consistent with those in the literature, and it was indicating that Neo promoted apoptosis in liver cancer cells and has significance for the study of anti-cancer mechanisms.

CONCLUSION

In conclusion, Neo showed inhibition effect on proliferation of HepG2 cells and SMMC-7721 cells *in vitro*. The survival rate of the two cells decreased and cell apoptosis rate increased in a dose-dependent and time-dependent way. The mechanism of antitumor effect may be related to cell cycle arresting in the G 0/G1 phase. The up-regulating expression of Bax, Caspase-3, Caspase-8 and Caspase-9 proteins and down-regulating the expression of Bcl-2 proteins in both cells may also show the mechanism of cell apoptosis when the dose of Neo is increasing. This study suggested that Neo may be a promising candidate or major compound for the

development of liver cancer, providing data support for further animal and clinical trials.

ACKNOWLEDGMENTS

This work was supported in part by the National Natural Science Foundation of China (NSFC 81208771), and Science and technology project of Chongqing Municipal Health bureau department (2021 MSXM101).

DATA AVAILABILITY STATEMENTS

The datasets generated during and/or analysed during the current study are available from the corresponding author on reasonable request.

REFERENCES

- Valery PC, Laversanne M, Clark PJ, et al. Projections of Primary Liver Cancer to 2030 in 30 Countries Worldwide. Hepatol. 2018; 67(2): 600-611.
- Firtina KZ, Koc D, Iscan E, et al. Elevated hepatocyte growth factor expression as an autocrine c-Met activation mechanism in acquired resistance to sorafenib in hepatocellular carcinom cells. J Cancer Sci 2016; 107: 407-416.
- Bray F, Ferlay J, Soerjomataram I, et al. Global Cancer Statistics 2018: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. A Cancer J Clinicians. 2018; 68(6): 394.424.
- 4. Fu J, Wang HY. Precision Diagnosis and Treatment of liver cancer in China. Cancer Letters. 2018; 412: 283-286.
- Li MF, Guan H, Zhang DD, et al. Effect of overexpression of PTEN on Apoptosis of liver cancer cells. Gene Mol Res. 2016;15(2).
- Taleghani A, Nasseri MA, Iranshahi M, et al. Synthesis of dual-action parthenolide prodrugs as potent anticancer agents. Bioorganic chem. 2017; 71: 128-134.
- Qiu Y, Wang JL. Research Progress on Anticancer Effects and Mechanisms of Three Natural Sesquiterpenoids. Journal of Jinzhou Medical University. 2020;41 (2): 109-112.
- Yu X, Zhang Q, Tian L, et al. Germacrane-type sesquiterpenoids with antiproliferative activities from Eupatorium chinense. J Nat Product. 2017; 81(1): 85.
- Abdel-Lateff A, Alarif W M, Ayyad SN, et al. New cytotoxic isoprenoid derivatives from the Red Sea soft coral Sarcophyton glaucum. J Nat Product Res. 2014; 29(1): 24.
- Fu J, Li FH, Li CK, et al. Reviews on natural monocyclic sesquiterpenoids and their bioactivities J. China J Chinese Materia Medica. 2019;44(17): 3672-3683.
- Qin RX, Zuo Q, Huang XH, et al. A new nor-sesquiterpene from Cassia occidentalis and its bioactivity. China J Chinese Materia Medica. 2016; 41(23): 4389-4392.
- Thao N P, Luyen B T T, Tai B H, et al. Chemical constituents of Miliusa balansae leaves and inhibition of nitric oxide production in lipopolysaccharide-induced RAW 264. 7 cells. Bioorganic Med Chem Letters. 2015; 25(18): 3859.
- Liu L, Liu R, Basnet BB, et al. New phenolic bisabolane sesquiterpenoid derivatives with cytotoxicity from Aspergillus tennes seensis. J Antibiotics. 2018; 71(5): 538.
- Dong X, Wang G, Zhuo Z, et al. Terpenoids from Ainsliaea latifolia and their cytotoxic activities. J Asian Nat Product Res. 2015;18(3): 232

- Lin X, Peng Z, Fu X, et al. Volatile oil from Saussurea lappa exerts antitumor efficacy by inhibiting epithelial growth factor receptor tyrosine kinase-mediated signaling pathway in hepatocellular carcinoma. Oncotarget. 2016; 48(2): 79761-79773.
- Kang Y, Wei L, Wang YX, et al. Effects of Rosiglitazone on cell cycle and apoptosis of Human Hepatoman Cell Line HepG2. Oncotarget. 2018; 33(4): 534-536.
- Hu Q, Jiang JH, Li Y, et al. Effects of Drug-containing Serum of Clerodendrum Bunge on Cell Cycle and Apoptosis of Human Hepatocellular Carcinoma MHCC97-H. J Hunan University of Chinese Med. 2019; 39(2): 1444-1448.
- Weinlich R, Oberst A, Beere A, et al. Necroptosis in development, inflammation and diseas. Nat rev mol cell biol. 2017; 18(2):127-136.
- Li F, Jin D, Tang C, et al. CEP55 promotes cell proliferation and inhibits apoptosis via the PI3 K/Akt/p21 signaling pathway in human glioma U251 cells. Oncology Letters. 2018;15(4): 4789-4796.
- Fazekas B, Carty MP, Nemeth I, et al. Hu COP1 contributes to the regulation of DNA repair in keratinocytes Molecular and Cellalar Biochenistry. 2017; 427(12): 103-109.
- Gross A. Bcl-2 family proteins as regulators of mitochondria metabolism. Biochimica et biophysica acta-molecular and cell biology of lipids. 2016; 1857(1): 1243-1246.
- Li H, Lv B, Kong L, et al. Mediates resistance of rat pheochromocytoma cells to hypoxia-induced apoptosis via the Bax/ Bcl-2/caspase-3 pathway. Int J Mol. 2017; 40(4): 1125-1133.
- Hengartner MO. The biochemistry of apoptosis. Nature. 2000; 407(6805): 770-776.
- 24. Wurstle ML, Laussmann MA, Rehm M, et al. The central role of initiator caspase-9 in apoptosis signal transduction and the regulation of its activation and activity on the apoptosome. Experimental Cell Res. 2012; 318: 1213-1220.
- 25. Araveti PB, Srivastava A. Curcumin induced oxidative stress causes autophagy and apoptosis in bovine leucocytes transformed by Theileria annulata. Cell Death Discovery. 2019; 5: 100
- Boice A, Bouchier-Hayes L. Targeting apoptotic caspases in cancer [J]. Biochimica et Biophysica Acta-Mol Cell Res. 2020;1867(6):1186-1188.
- Ko HM, Joo SH, Jo JH, et al. Liver-Wrapping, Nitric Oxide-Releasing Nanofiber Downregulates Cleaved Caspase-3 and Bax Expression on Rat Hepatic Ischemia-Reperfusion Injury. Transplantation Proceedings. 2017; 49(5): 1170-1174.
- 28. Hua PY, Sun M, Zhang GX, et al. Costunolide induces apoptosis through generation of ROS and activation of P53 in human esophageal cancer Eca-109 cells. J Biochem Mol Toxicol. 2016; 30(9): 462-469.
- Guo F, Li LL, Zang L, et al. Study of mechanism of curcumol on mediating anti breast cancer via down-regulation the expression of Bcl-2 protein. The Chinese J Clin Pharmacol. 2018; 10: 1175-1178.
- Marquardt JU, Edlich F. Predisposition to Apoptosis in Hepatocellular Carcinoma: From Mechanistic Insights to Therapeutic Strategies. Frontiers in Oncol. 2019; 9: 1421.