

Study of the Effect of Catechin of Green Tea Extract upon Sputum Smear Conversion Rate , Serum Malondialdehyde and Blood Iron Indices in Tuberculosis Patients

Honarvar.M.R¹ Eghtesadi S², Gill.P³, Jazayeri.S⁴, Vakili M.⁵, Shamsardakani.M.R⁶, Abbasi.A⁷, Eghtesadi M⁸

ABSTRACT: Green tea with possessing iron chelating properties can be useful in TB treatment and management. We studied the effect of green tea consumption on iron status and improving process of pulmonary tuberculosis treatment (accelerating the negative sputum smear, reducing the level of oxidative stress). Following the approval by Ethics Committee for Human Studies of Golestan and Tehran Universities of Medical Sciences and also obtaining the written consent of patients , this double-blinded randomized clinical trial study, was conducted on patients with TB, who were assigned randomly to the intervention group (41 patients) receiving 500 mg catechin of green tea extract and the control group (39 subjects) receiving placebo for two months, since the beginning of concomitant anti-TB treatment . Sputum evaluation was carried out on three slides using the Ziehl Nelson method. At first, the demographic and dietary intake data were obtained. . After obtaining 10 ml of venous blood, Hemoglobin (Hb), Transferrin, Ferritin, Total iron binding capacity (TIBC), Iron and Serum malondialdehyde (MDA) were measured at the beginning and end of the study. Sputum samples were collected from the third week (every 10 days) and the reduction of microbial load was also tested until sputum smear became negative. Data were processed using independent and paired t-test, McNemar, Wilcoxon, Kaplan-Meier, Log-rank test and Cox regression model. Pvalue was taken significant as <0.05. Average daily energy intake of patients was 1518±431 kcal, distribution of which was as follow: carbohydrates (58%), protein (17%) and fat (22%). Vitamin D and Zinc intake of patients were less and iron intake was higher than the DRI. Weight changes in both groups of placebo and green tea had tendency of increase with a significant difference at two and six month follow ups (p<0.0001). However, there were no significant changes due to intervention compared to placebo. Sputum

conversion time (days) was 52.5± 24.5 (median= 53 days) and 40.6 ± 22.5 (median= 29 days) in placebo and catechin groups, respectively. The proportion of patients in the green tea group based on criterion of ; the short duration of being negative sputum smear; was significantly higher than the placebo group (p=0.032). To measure the mean of iron status after intervention, ANCOVA test showed mean difference level (Pvalue) in both groups for Hb, iron, TIBC, transferrin and ferritin as of: 0.004, 0.56, 0.65, 0.38 and 0.16, respectively which means that increase of hemoglobin in the green tea group was significant compared with the placebo group. There was just a 9.2 nmol/ml difference between the two groups for MDA at the beginning of study, which was not statistically significant (p=0.078) whereas, it was increased to 24.8 nmol/ml after the intervention, indicating a significant difference (p<0.001). The decline value was estimated -45.45 ± 14.69 nmol/ml for catechin group and -19.91 ± 18.38 nmol/ml for placebo group. In conclusion green tea can systematically reduce the inflammatory elements and oxidants (decrease of MDA as fatty acids oxidation indicator), and consequently, can improve the hematopoiesis and hemoglobin level. Therefore, localized inflammation and damage in the lung is reduced, and adjunct to antimicrobial therapy, accelerate sputum smear conversion, disease amelioration and treatment improvement. Finally, given the higher iron intake despite of lower micronutrients and macronutrients in diet of our patients, and considering the iron effect on mycobacterium survival and the incidence and exacerbation of inflammatory complications in patients, it seems that policy of mandatory flour fortification with iron, especially in provinces such as Golestan, must be viewed cautiously and its further implementation being revised meticulously. Email: segtesadi@gmail.comes..

Keywords: Sputum Smear, Weight, Blood Indices , Green Tea, Pulmonary Tuberculosis.

INTRODUCTION

Supportive studies about improving anti-tuberculosis treatment, indicate that special nutritional adjunct therapies, play an important role in treatment of TB or its side effects(1-15). High prevalence of TB in Golestan Province(near Caspian Sea) of Iran motivated us to use supplementation with nutritional elements such as green tea extract to promote healing and decrease secondary complications of this disease(16,19-22). Iron is basically essential for Mycobacterium tuberculosis replication within the host and its pathogenicity(24-25,28-29). Green tea with possessing iron chelating properties and its effect on gene expression of antioxidant proteins(Hepcidin and Transferrin) involved in regulating intracellular iron can be useful in TB treatment and management(30-36). Tea consumption is a part of food culture of Iranian people as a popular social entertainment drink, especially in the Golestan province. It may be useful to encourage families to drink green tea if its benefits get proven scientifically.

Polyphenols fight inflammation through inhibition of the synthesis and function of inflammatory mediators like eicosanoids and cytokines .

The increase in the level of inflammatory cytokines of the plasma may be associated with malnutrition in active pulmonary TB . Oxidative stress may suppress the immune system(16-17). In patients with TB, consumption of antioxidant supplements may tackle oxidative stress(26). Green tea can protect the immune system against oxidant compounds and free radicals through controlling oxidative stress and boost its function as well . It seems that green tea may help to accelerate the process of recovery and weight gain in TB patients through decreasing oxidative stress(30-36).

Conversion of the sputum smear after two months of intensive antibiotic therapy in sputum smear positive patients is an important indicator of treatment(18) . Acceleration in sputum smear conversion helps with faster improvement and decreased probability of the transfer of TB in smear-positive patients with tuberculosis(26-27). Green tea catechin is an important potential immunotherapeutic agent against respiratory infections and negatively affects the survival of M. tuberculosis through inhibition of the NF- κ B pathway in vitro (13,19,37-40). For this reason, consumption of green tea or supplementation with concentrated polyphenols derived from green tea can be regarded as an adjuvant therapy in patients receiving antibiotics(41-44) .

¹School of Public Health ,Golestan University of Medical Sciences, Gorgan, Iran ²Azad University , Science and Research Branch,School of Medical Sciences and Technology, Tehran, Iran ³Departments of PhysioPharmacology&NanoBioMedicine,Research Center for Immunogenetics, Faculty of Medicine, Mazandaran University of Medical Sciences ,Sari, Iran.⁴Department of Nutrition, School of Public Health, Iran University of Medical Sciences, Tehran, Iran.,⁵Department of Health and Social Medicine, Health Management and Social Development Research Center, Golestan University of Medical Sciences, Gorgan, Iran.⁶Department of Pharmacognosy, Faculty of Pharmacy and Medicinal Plants Research Center , Tehran University of Medical Sciences , Tehran , Iran.⁷Department of Infection, Faculty of Medicine, Golestan University of Medical Sciences, Gorgan, Iran,⁸Azad University,School of Medical Sciences, Tehran Branch, Department of Research, Tehran, Iran

Correspondence Author: Shahryar Eghtesadi, Azad University , Science and Research Branch,School of Medical Sciences and Technology, Tehran, Iran. emai: segtesadi@gmail.com



This open-access article is distributed under the terms of the Creative Commons Attribution Non-Commercial License (CC BY-NC) (<http://creativecommons.org/licenses/by-nc/4.0/>), which permits reuse, distribution and reproduction of the article, provided that the original work is properly cited and the reuse is restricted to noncommercial purposes. For commercial reuse, contact reprints@pulsus.com

Considering the lack of such studies in Iran, we decided to perform this study to evaluate the effect of green tea as an adjuvant therapy on shortening the sputum smear conversion time, weight and its effect upon the malondialdehyde and blood iron indices in patients with smear positive pulmonary tuberculosis. The primary outcome of the study was the sputum smear conversion time and the secondary outcome was changes in weight and BMI within two months.

Materials and Methods:

This double-blinded randomized clinical trial study, was conducted on patients with TB, who were assigned randomly to the green tea group (41 patients) receiving 500 mg catechin of green tea extract and the control group (39 subjects) receiving placebo for two months since the beginning of concomitant anti-TB treatment(Figure1)(20-21).This study included patients with smear positive pulmonary TB, who were diagnosed with this disease, based on clinical manifestations and sputum smear positivity according to the guidelines of the Iranian Ministry of Health. The patients received Isoniazide, Rifampin, Pyrazinamide, and Ethambutol according to the DOTS(Directly Observed Treatment, Short Course)strategy.In this protocol the patients receive a 4-drug regimen at the first 2month, with a two-drug regimen in the following four month of the treatment. Sputum smear evaluation was performed at two and six month post-treatment to assess improvement. Random allocation and allocation concealment was observed. Height and weight were measured at first and two and six months thereafter. From day 20 onward, sputum smears were obtained every 10 days and the reduction of microbial load was also tested until sputum smear became negative.Sputum evaluations were performed on three slides using the Ziehl Nelsen method, and sputum smear conversion was defined as two negative slides for M.tuberculosis.Survival time(in months) was calculated from the date of diagnosis to the date of death or last follow-up. Failure was defined as death by any cause during the follow-up period, and the sputum smear conversion time of more than two months was censored in the patients. The duration of intervention was considered from the commencement of the therapy to the date of sputum conversion in the second month of the treatment(41-44).

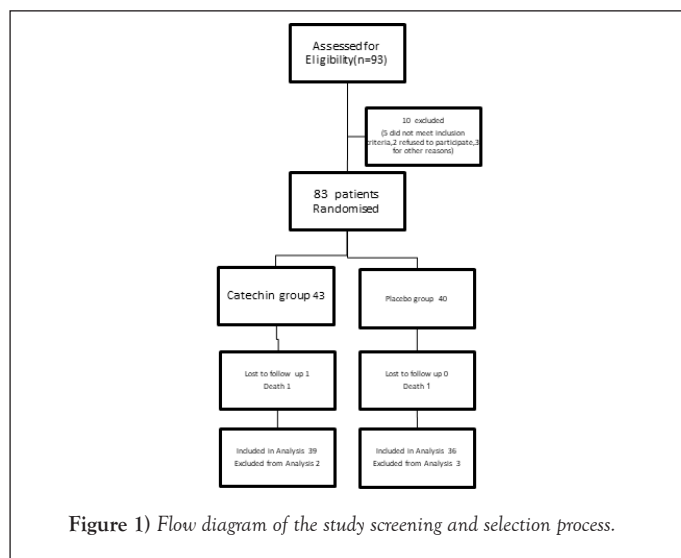


Figure 1) Flow diagram of the study screening and selection process.

Demographic, food frequency questionnaires and 24-hour dietary recalls of three non-consecutive days were completed. After obtaining 10 ml of venous blood, Hemoglobin (Hb), Transferrin, Ferritin, Total iron binding capacity (TIBC), Iron and Serum malondialdehyde (MDA) were measured at the beginning and the end of the study. Data were processed using independent and paired t-test, McNemar, Wilcoxon, Kaplan-Meier, Log-rank test, Cox regression model and nutrition4 software. P values < 0.05 were considered significant.

Results and Discussion:

At the beginning of the survey, two groups were similar according to age, gender and clinical status (p>0.05)(Table1).Average daily energy intake of patients was 1518± 434 kcal, distribution of which was as follow: carbohydrates (58%), protein (17%) and fat (22%).Vitamin D and Zinc intake of patients

were less and iron intake was higher than the DRI

TABLE 1

Comparison of some selected nutrient intakes between Placebo and Catechin groups of TB subjects

Nutrients	Placebo	Catechin	P value
Energy(Kcal/day)	1499±15	1531±453	0.751
Carbohydrate(g/day)	221±74	219±80	0.903
Protein(g/day)	62.8±18.4	67.7±23.3	0.312
Fat(g/day)	42±22	43±18	0.723
Vitamin D(µg/day)	1.6±2.3	1.5±1.8	0.935
VitaminA(µg/day)	842±1364	852±2044	0.972
VitaminE(µg/day)	3.5±5	2.6±4.2	0.427
Fe(mg/day)	17.3±9.8	9.5±4.1	0.322
Zn(mg/day)	5.72.5	6.7±3.2	0.151

Weight changes in both groups of placebo and green tea had tendency of increase with a significant difference at two and six month follow ups (p<0.0001). However, there were no significant changes due to intervention compared to placebo(Figure2)

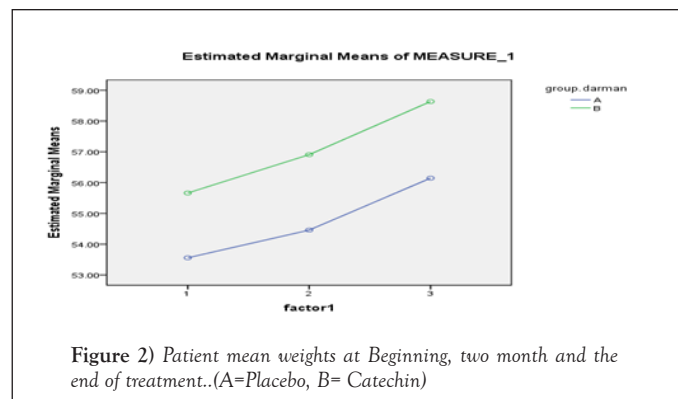


Figure 2) Patient mean weights at Beginning, two month and the end of treatment..(A=Placebo, B= Catechin)

Sputum conversion time (days) was 52.5± 24.5 (median= 53 days) and 40.6 ± 22.5 (median= 29 days) in placebo and catechin groups, respectively. The proportion of patients in the green tea group based on criterion of ; the short duration of being negative sputum smear; was significantly higher than the placebo group, Log Rank(Mantel-cox)=4.61, (p=0.032). (Figure3)

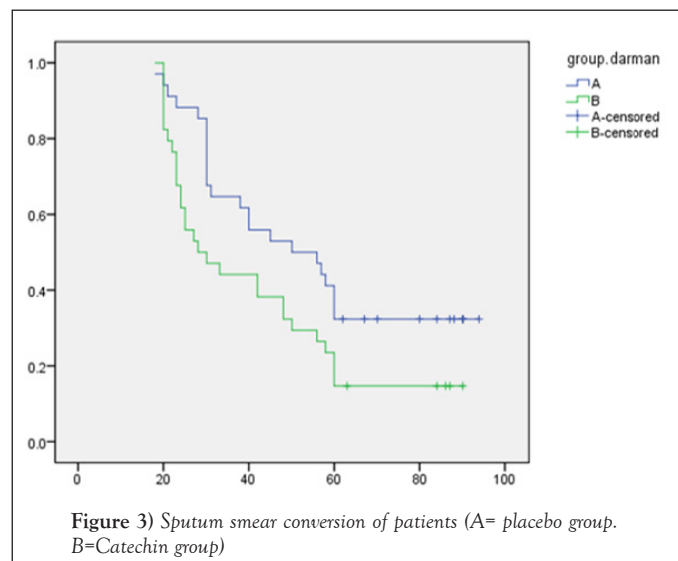
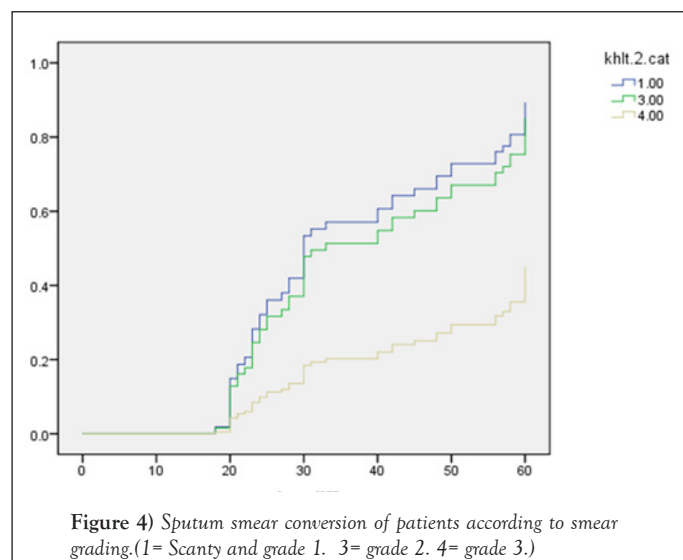


Figure 3) Sputum smear conversion of patients (A= placebo group. B=Catechin group)

Based on the Cox regression model, the hazard ratio of the relative risk of delay in sputum smear conversion was 3.7 (CI 1.7-8.6, p=0.002) in the higher microbial load group compared to the placebo group and 0.54 (CI 0.31-0.94) in the intervention group compared to the placebo group(Figure4)

Considering the status of the sputum microbial load, the time to sputum smear conversion is expected to be shorter in the scanty and +1 group versus the +3 group. The two groups of GT and placebo were compared for days to sputum smear conversion considering the microbial load of the sputum at the beginning of the study. (Figure4).Therefore ,it was found that the sputum smear conversion time was significantly shorter in the GT group than the placebo group (p=0.031) (Figure3),and the risk of delay in sputum conversion augmented with the increase in the microbial load and decreased with the consumption of green tea(Figure4).



To measure the mean of iron status after intervention,ANCOVA test showed mean difference level (P value) in both groups for Hb, iron, TIBC, transferrin and ferritin as of: 0.004, 0.56, 0.65, 0.38 and 0.16, respectively which means that increase of hemoglobin in the green tea group was significant.

compared to the placebo group. There was just a 9.2 nmol/ml difference between the two groups for MDA at the beginning of the study, which was not statistically significant (p=0.078) whereas, it was increased to 24.8 nmol/ml after the intervention, indicating a significant difference (p<0.001). The decline value was estimated -45.45 ± 14.69 nmol/ml for catechin group and -19.91 ± 18.38 nmol/ml for placebo group.

To our knowledge ,our study might be the first to be conducted on the use of green tea extract as a supplement in the treatment of tuberculosis in Iran.The present survey showed the Green tea can systematically reduce the inflammatory elements and oxidants (decrease of MDA as fatty acids oxidation indicator), and consequently, can improve the hematopoiesis and hemoglobin level. Therefore, localized inflammation and damage in the lung is reduced, and adjunct to antimicrobial therapy, accelerate sputum smear conversion, disease amelioration and treatment improvement. If researchers implement catechin as an adjuvant with antibiotic therapy as well as

nutritional modification especially effective micronutrients on the immune status of patients, more improvement of disease treatment might be achieved.

Our study well documented the effect of catechin on sputum smear conversion. The microbial load of the sputum decreased faster in patients who received catechin compared to the control group.In fact high grade of smear positivity was associated with failure in sputum smear conversion, due to the higher initial bacterial load that required a longer time for clearance in the early stages of treatment.In our study a higher microbial load was correlated with delay in sputum smear conversion.It seems that the contribution of green tea to decreasing the inflammatory status of the patients has a little effect on improving the nutritional status of the patients when compared to the effect of antibacterial therapy. Some studies have evaluated the status of the inflammatory indices. For example, Kim et al reported that the levels of TNF α and IL-6 were higher in malnourished TB patients than the control group and well-nourished TB patients (31) and the level of C-reactive protein (CRP) and the proportion of CRP to albumin was

lower in patients not afflicted with TB than TB patients 30 and 60 days after treatment (32).

Numerous studies have highlighted the role of green tea, especially its catechins, in regulating inflammatory reactions (21, 33-35). Green tea entraps reactive oxygen species (ROS) and is protective against oxidative stress. Agarwal et al showed the effect of green tea extract on the reduction of oxidative stress in patients receiving anti TB treatment (23).

In short,in our study green tea had a little effect on weight gain but because its catechins contents are natural compounds with relatively safe profiles, the use of green tea as an adjuvant therapy in TB patients may be a new method for faster rehabilitation of the patients through shortening the time to sputum smear conversion.

The main limitation of our study was the lack of sputum culture, which is considered the gold standard of TB diagnosis. In addition, difficulty obtaining sputum specimens during treatment, due to the clearance of the patients'lungs,was another limitation of the study. Finally , considering the DOTS protocol,few patients were asked to take X-ray to follow the treatment.

It is important to mention that given the higher iron intake despite of lower micronutrients and macronutrients in diet of our patients, and considering the iron effect on mycobacterium survival and the incidence and exacerbation of inflammatory complications in patients, it seems that policy of mandatory flour fortification with iron,especially in provinces such as Golestan, must be viewed cautiously and its further implementation being revised meticulously

REFERENCES

1. Ministry of Health and Medical Education of Iran. Communicable disease control center. Department of Tuberculosis and Leprosy [Internet]. [cited 2014 Dec 11]. Available from: <http://tb-lep.behdasht.gov.ir/Default.aspx>
2. PrayGod G, Range N, Faurholt-Jepsen D, Jeremiah K, Faurholt-Jepsen M, Aabye MG, et al. Daily Multi-Micronutrient Supplementation during Tuberculosis Treatment Increases Weight and Grip Strength among HIV-Uninfected but Not HIV-Infected Patients in Mwanza, Tanzania. *J Nutr.* 2011 Apr 1;141(4):685-91.
3. Schön T, Idh J, Westman A, Elias D, Abate E, Diro E, et al. Effects of a food supplement rich in arginine in patients with smear positive pulmonary tuberculosis - A randomised trial. *Tuberculosis.* 2011 Sep;91(5):370-7.
4. Jordao L, Lengeling A, Bordat Y, Boudou F, Gicquel B, Neyrolles O, et al. Effects of omega-3 and -6 fatty acids on Mycobacterium tuberculosis in macrophages and in mice. *Microbes Infect.* 2008 Oct;10(12-13):1379-86.
5. Crewe RM. HIV/AIDS ,TB and Nutrition [Internet]. Available from: <http://www.assaf.org.za>
6. Papatthakis P, Piwoz E. Nutrition and Tuberculosis: A Review of the Literature and Considerations for TB Control Programs. *U S Agency Int Dev Afr Health* 2010 Proj. 2008;1.
7. Abba K, Sudarsanam TD, Grobler L, Volmink J. Nutritional supplements for people being treated for active tuberculosis. *Cochrane Database Syst Rev Online.* 2008;(4):CD006086.
8. Sudarsanam TD, John J, Kang G, Mahendri V, Gerrior J, Franciosa M, et al. Pilot randomized trial of nutritional supplementation in patients with tuberculosis and HIV-tuberculosis coinfection receiving directly observed short-course chemotherapy for tuberculosis. *Trop Med Int Health TM IH.* 2011 Jun;16(6):699-706.
9. Jahnavi G,, Sudha C H. Randomised controlled trial of food supplements in patients with newly diagnosed tuberculosis and wasting. *Singap Med J* 2010 5112 957-962.
10. Range N, Andersen ÅB, Magnussen P, Mugomela A, Friis H. The effect of micronutrient supplementation on treatment outcome in patients with pulmonary tuberculosis: a randomized controlled trial in Mwanza, Tanzania. *Trop Med Int Health.* 2005 Sep 1;10(9):826-32.
11. Visser ME, Grewal HM, Swart EC, Dhansay MA, Walzl G, Swanevelder S, et al. The effect of vitamin A and zinc supplementation on treatment

- outcomes in pulmonary tuberculosis: a randomized controlled trial. *Am J Clin Nutr.* 2011 Jan 1;93(1):93-100.
12. Hegde ML, Bharathi P, Suram A, Venugopal C, Jagannathan R, Poddar P, et al. Challenges Associated with Metal Chelation Therapy in Alzheimer's Disease. *J Alzheimers Dis JAD.* 2009 Jul;17(3):457-68.
 13. Anand PK, Kaul D, Sharma M. Green tea polyphenol inhibits *Mycobacterium tuberculosis* survival within human macrophages. *Int J Biochem Cell Biol.* 2006;38(4):600-9.
 14. Serafini M, Peluso I, Raguzzini A. Flavonoids as anti-inflammatory agents. *Proc Nutr Soc.* 2010 Aug;69(03):273-8.
 15. Kim JH, Lee C-T, Yoon HI, Song J, Shin WG, Lee JH. Relation of ghrelin, leptin and inflammatory markers to nutritional status in active pulmonary tuberculosis. *Clin Nutr.* 2010 Aug;29(4):512-8.
 16. Matsumoto K, Yamada H, Takuma N, Niino H, Sagesaka YM. Effects of Green Tea Catechins and Theanine on Preventing Influenza Infection among Healthcare Workers: A Randomized Controlled Trial. *BMC Complement Altern Med.* 2011 Feb 21;11:15.
 17. Neyestani T, Khalaji N. Inhibitory effects of black tea (*Camellia sinensis*) extracts on *Streptococcus pyogenes*: A comparison between black and green teas in vitro. *Iran J Nutr Sci Food Technol.* 2007 Jun 15;2(1):41-7.
 18. Dept WHO. Compendium of indicators for monitoring and evaluating national tuberculosis programs. 2004 [cited 2014 Nov 6]; Available from: <http://apps.who.int/iris/handle/10665/68768>
 19. Yamamoto Y, Matsunaga K, Friedman H. Protective effects of green tea catechins on alveolar macrophages against bacterial infections. *Biofactors.* 2004;21(1-4):119-21.
 20. Fatima Z, Hameed S, Islam N. Epigallocatechin-3-gallate (EGCG), a green tea polyphenol suppresses bacilli-induced augmented expression of *Mycobacterium tuberculosis* 85B and proinflammatory TNF- α in human monocytes. *Int J Sci Res Publ.* 2012;90.
 21. Anand PK, Kaul D, Sharma M. Green tea polyphenol inhibits *Mycobacterium tuberculosis* survival within human macrophages. *Int J Biochem Cell Biol.* 2006;38(4):600-9.
 22. Agarwal A, Prasad R, Jain A. Effect of green tea extract (catechins) in reducing oxidative stress seen in patients of pulmonary tuberculosis on DOTS Cat I regimen. *Phytomedicine Int J Phytother Phytopharm.* 2010 Jan;17(1):23-7.
 23. PrayGod G, Range N, Faurholt-Jepsen D, Jeremiah K, Faurholt-Jepsen M, Aabye MG, et al. Weight, body composition and handgrip strength among pulmonary tuberculosis patients: a matched cross-sectional study in Mwanza, Tanzania. *Trans R Soc Trop Med Hyg.* 2011 Mar;105(3):140-7.
 24. Zachariah R, Spielmann MP, Harries AD, Salaniponi FML. Moderate to severe malnutrition in patients with tuberculosis is a risk factor associated with early death. *Trans R Soc Trop Med Hyg.* 2002 Jun 1;96(3):291-4.
 25. Schwenk A, Hodgson L, Wright A, Ward LC, Rayner CF, Grubnic S, et al. Nutrient partitioning during treatment of tuberculosis: gain in body fat mass but not in protein mass. *Am J Clin Nutr.* 2004 Jun 1;79(6):1006-12.
 26. Lee JH, Chang JH. Changes of plasma interleukin-1 receptor antagonist, interleukin-8 and other serologic markers during chemotherapy in patients with active pulmonary tuberculosis. *Korean J Intern Med.* 2003 Sep;18(3):138-45.
 27. Klausner JD, Makonkawkeyoon S, Akarasewi P, Nakata K, Kasinrek W, Corral L, et al. The effect of thalidomide on the pathogenesis of human immunodeficiency virus type 1 and *M. tuberculosis* infection. *JAIDS J Acquir Immune Defic Syndr.* 1996;11(3):247-57.
 28. Range N, Andersen AB, Magnussen P, Mugomela A, Friis H. The effect of micronutrient supplementation on treatment outcome in patients with pulmonary tuberculosis: a randomized controlled trial in Mwanza, Tanzania. *Trop Med Int Health TM IH.* 2005 Sep;10(9):826-32.
 29. Paton NI, Chua Y-K, Earnest A, Chee CB. Randomized controlled trial of nutritional supplementation in patients with newly diagnosed tuberculosis and wasting. *Am J Clin Nutr.* 2004 Aug 1;80(2):460-5.
 30. Karyadi E, West CE, Schultink W, Nelwan RH, Gross R, Amin Z, et al. A double-blind, placebo-controlled study of vitamin A and zinc supplementation in persons with tuberculosis in Indonesia: effects on clinical response and nutritional status. *Am J Clin Nutr.* 2002 Apr 1;75(4):720-7.
 31. Kim JH, Lee C-T, Yoon HI, Song J, Shin WG, Lee JH. Relation of ghrelin, leptin and inflammatory markers to nutritional status in active pulmonary tuberculosis. *Clin Nutr.* 2010 Aug;29(4):512-8.
 32. Moraes ML de, Ramalho DM de P, Delogo KN, Miranda PFC, Mesquita EDD, Oliveira HM de MG de, et al. Association of Serum Levels of Iron, Copper, and Zinc, and Inflammatory Markers with Bacteriological Sputum Conversion During Tuberculosis Treatment. *Biol Trace Elem Res.* 2014 Aug 1;160(2):176-84.
 33. Von Staszewski M, Pilosof AMR, Jagus RJ. Antioxidant and antimicrobial performance of different Argentinean green tea varieties as affected by whey proteins. *Food Chem.* 2011 Mar 1;125(1):186-92.
 34. Singh BN, Shankar S, Srivastava RK. Green tea catechin, epigallocatechin-3-gallate (EGCG): Mechanisms, perspectives and clinical applications. *Biochem Pharmacol.* 2011 Dec 15;82(12):1807-21.
 35. Basu A, Betts NM, Mulugeta A, Tong C, Newman E, Lyons TJ. Green tea supplementation increases glutathione and plasma antioxidant capacity in adults with the metabolic syndrome. *Nutr Res.* 2013 Mar;33(3):180-7.
 36. Martins N, Morris P, Kelly PM. Food incentives to improve completion of tuberculosis treatment: randomised controlled trial in Dili, Timor-Leste. *BMJ.* 2009 Oct 26;339(oct26 1):b4248-b4248.
 37. Schön T, Elias D, Moges F, Melese E, Tessema T, Stendahl O, et al. Arginine as an adjuvant to chemotherapy improves clinical outcome in active tuberculosis. *Eur Respir J.* 2003 Mar 1;21(3):483-8.
 38. Jahnvi G, Sudha CH. Randomised controlled trial of food supplements in patients with newly diagnosed tuberculosis and wasting. *Singapore Med J.* 2010 Dec;51(12):957-62.
 39. Coussens AK, Wilkinson RJ, Hanifa Y, Nikolayevskyy V, Elkington PT, Islam K, et al. Vitamin D accelerates resolution of inflammatory responses during tuberculosis treatment. *Proc Natl Acad Sci.* 2012 Sep 18;109(38):15449-54.
 40. Visser ME, Grewal HM, Swart EC, Dhansay MA, Walzl G, Swanevelder S, et al. The effect of vitamin A and zinc supplementation on treatment outcomes in pulmonary tuberculosis: a randomized controlled trial. *Am J Clin Nutr.* 2011 Jan 1;93(1):93-100.
 41. Jayakody W, Harries AD, Malhotra S, de Alwis S, Samaraweera S, Pallewatta N. Characteristics and outcomes of tuberculosis patients who fail to smear convert at two months in Sri Lanka. *Public Health Action.* 2013 Mar 21;3(1):26-30.
 42. Dominguez-Castellano A, Muniain MA, Rodriguez-Baño J, Garcia M, Rios MJ, Galvez J, et al. Factors associated with time to sputum smear conversion in active pulmonary tuberculosis. *Int J Tuberc Lung Dis.* 2003 May 1;7(5):432-8.
 43. Singla R, Osman M. M, Khan N, Al-Sharif N, Al-Sayegh M. O, Shaikh M. A. Factors predicting persistent sputum smear positivity among pulmonary tuberculosis patients 2 months after treatment. *Int J Tuberc Lung Dis.* 2003 Jan 1;7(1):58-64.
 44. Parikh R, Nataraj G, Kanade S, Khatri V, Mehta P. Time to sputum conversion in smear positive pulmonary TB patients on category I DOTS and factors delaying it. *J Assoc Physicians India.* 2012;60:22-6.